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1 A new model of ozone stress in wheat including grain yield loss and plant 2 acclimation to the pollutant

3 I. Droutsas^{a,b,*}, A. J. Challinor^{a,b}, S. R. Arnold^{a,b}, T. N. Mikkelsen^c, E. M. Ø. Hansen^d

4 ^a*Institute for Climate and Atmospheric Science, School of Earth and Environment, University of Leeds, LS2 9JT Leeds, UK*

5 ^b*Priestley International Centre for Climate, University of Leeds, LS2 9JT, Leeds, UK*

6 ^c*Department of Environmental Engineering, Technical University of Denmark, DK-2800 Lyngby, Denmark*

7 ^d*Department of People and Technology, Roskilde University, Universitetsvej 1, DK-4000 Roskilde, Denmark*

8 Abstract

9 Surface ozone (O₃) is an important air pollutant globally and enhanced concentrations lead to crop yield
10 penalties in many parts of the world. Crop models simulate production and yield and they are often used
11 for various applications. However, most of the existing models neglect the effect of O₃ and only limited
12 parameterization schemes exist. In addition, the existing O₃ modelling approaches do not take into account
13 the plant acclimation to the pollutant as a mechanism of survival and maintenance of performance. Here,
14 we introduce a simple modelling method to simulate the O₃ damage to wheat with consideration of the plant
15 acclimation process. The O₃ parameterization scheme was incorporated into the GLAM-Parti crop model,
16 resulting in a new model version GLAM-ROC (i.e. GLAM - Relative Ozone Concentrations). The new model
17 simulates the effect of O₃ on crop growth and development and was evaluated against data from control-
18 environment chambers with high O₃ concentration levels and variable duration of exposure to the pollutant.
19 GLAM-ROC successfully reproduced the observed plant response to O₃ as well as the final biomass and
20 yield. The incorporation of plant acclimation allowed the prediction of crop yield loss at variable duration of O₃
21 exposure. The statistical response formula neglected the acclimation process and overestimated the relative
22 O₃ damage to yield by 56.5%, when fumigation increased from 32 to 106 days. We conclude that the plant
23 acclimation to chronic O₃ environment is significant and should be taken into account for the effect of O₃ on
24 wheat performance and yield.

25 *Keywords:* Crop model, Ozone, Wheat, Acclimation, GLAM-ROC, GLAM-PARTI

26 1. Introduction

27 Ground-level ozone (O₃) is a highly phytotoxic air pollutant at global scale (Ashmore, 2005; Ainsworth,
28 2017). Current O₃ levels induce crop yield damage and lead to decreased food supply and economic loss
29 (Emberson et al., 2009; McGrath et al., 2015). Avnery et al. (2011) estimated that global yields of soybean and
30 wheat were reduced by up to 14 and 15% respectively for the year 2000 due to O₃ pollution. Mills et al. (2018)

31 estimated that in highly polluted regions of N India and NW China the O₃ damage to wheat yield exceeded
32 15% on average for the years 2010 - 2012. O₃ concentrations are projected to remain enhanced in many
33 regions in the future, potentially posing serious threat to agriculture (Sicard et al., 2017).

34 The main mechanisms through which O₃ affects crops are by inhibiting photosynthesis, accelerating the
35 plant senescence rate and causing leaf chlorosis or necrosis under acute exposure (Heath, 1994;
36 Farage and Long, 1999; Fiscus et al., 2005). These effects result in decreased photosynthate allocation to
37 the grain, reduced productivity and lower yield (Wilkinson et al., 2012). The range of effects depends upon the
38 concentration level of the pollutant, the time and duration of exposure (Heath et al., 2009), the plant sensitivity
39 (Van Goethem et al., 2013) and the stage of plant development (Tiedemann and Pfähler, 1994; Mulholland
40 et al., 1998).

41 The effect of O₃ on crop yield has been extensively studied and various modelling approaches have been
42 suggested. Initially, different metrics were developed to link the plant O₃ exposure to the reduction in grain
43 yield. These metrics accumulate the O₃ concentration during the crop-growing season (e.g. AOT40, M7,
44 SUM06, W126) and relate the effect to yield according to a statistical response function (e.g. Fuhrer et al.,
45 1997; Mauzerall and Wang, 2001). However, various interactions between the crops and their surrounding
46 environment modify the magnitude of this relationship (Musselman et al., 2006). This is a major limitation
47 of exposure-based approaches and so O₃ effects were later introduced into more complex models of plant
48 growth and development.

49 The family of more complex models tend to use stomatal flux parameterizations (e.g. Emberson et al.,
50 2000; Pleijel et al., 2007) such as those found in crop models (e.g. Ewert and Porter, 2000; Schauburger et al.,
51 2019). These models can improve upon the exposure-based estimation of O₃ damage to crop productivity
52 and yield by simulating the stomatal limitations which regulate O₃ uptake by the plants (Challinor et al., 2009;
53 Pleijel et al., 2004). Nevertheless, the modelling of stomatal conductance is difficult and it is not clear which
54 of the many models of different complexity (Damour et al., 2010) is closest to reality. Modelled responses
55 to CO₂ concentration, temperature, air humidity, light and soil water content differ (Buckley and Mott, 2013),
56 resulting in different errors in the calculation of O₃ uptake and damage.

57 Plants can adjust their physiological and metabolic processes to enhance their stress tolerance over time
58 (Bruce et al., 2007). Under long-term O₃ exposure, the plant anti-oxidative enzyme activity increases (Gille-
59 spie et al., 2011, 2012), working as a mechanism of defence in favour of closing stomata to avoid take-up of
60 O₃ (Feng et al., 2016). This acclimation mechanism allows stomata to remain partially open and support gas

61 exchange for photosynthesis, thus avoiding high reductions in biomass accumulation (Chen et al., 2011). The
62 acclimation process in stress environments improves the plant response to the stressor (Kollist et al., 2018)
63 and leads to optimisation of productivity and yield. Held et al. (1991) exposed radish plants to high O₃ either
64 six days after germination or three days later and found that the plants which were exposed to the the pollutant
65 for the longer period exhibited higher dry mass than the plants exposed to O₃ later, implying an acclimation
66 mechanism. Trees can also compensate for the negative O₃ effects by activating acclimation mechanisms.
67 Mikkelsen and Ro-Poulsen (1994) reported higher photosynthesis levels of Norway spruce in the morning
68 before 8-h daily O₃ fumigation, as well as five days post-O₃ fumigation in comparison with non-fumigated
69 trees. Crop models do not usually parameterize for plant acclimation to chronic O₃ stress. One barrier to
70 the development of acclimation parameterizations in crop models is that the models are not evaluated under
71 variable duration of exposure to O₃.

72 The purpose of this study is to incorporate the effect of O₃ into a crop model by accounting for the concen-
73 tration level of the pollutant, the stage of plant development and the duration of plant exposure. The wheat
74 crop was selected as case study since it is particularly sensitive to O₃ (Barnes et al., 1990; Farage et al.,
75 1991; Burney and Ramanathan, 2014), an important staple crop at global level (FAO et al., 2017) and there
76 is excellent data availability. The GLAM-Parti crop model was used to incorporate the effect of O₃ on wheat,
77 resulting in a new model version called GLAM-ROC (i.e. GLAM-Relative Ozone Concentrations). Prior to
78 the incorporation of the O₃ effect, the allometric relationships for partitioning plant biomass in GLAM-Parti
79 were extended to the full crop cycle, since the model was previously developed with the GLAM approach for
80 post-anthesis crop growth and development.

81 **2. Materials and Methods**

82 *2.1. Wheat varieties and growing conditions*

83 Two modern spring wheat varieties were considered in this study, Lennox (Saaten-Union) used in south-
84 ern France and KWS Bittern (DanishAgro) used in Denmark. Lennox was used for the development of the O₃
85 algorithm in the model and KWS Bittern for the model evaluation. The plants were grown in 24 m² chambers
86 in the RERAF (Risoe Environmental Risk Assessment Facility) climate phytotron at the Technical University of
87 Denmark, Campus Risø, Roskilde. The plants were sown in 11 L pots filled with 4 kg of sphagnum (Pindstrup
88 Substrate No. 4, Pindstrup Mosebrug A/S, Ryomgaard, Denmark) and reduced to eight plants after germina-
89 tion, corresponding to ~ 165 plants m⁻². Light intensity in the chambers was approximately 400 mol photons

90 $\text{m}^{-2} \text{s}^{-1}$ PAR (photosynthetically active radiation) at canopy height and was provided for 16 h d^{-1} . The growing
 91 conditions in the chambers are shown in Table 1. The plants were watered three times a week to ensure
 92 full water supply. No additional nutrients were added to the pots since the sphagnum was nutrient enriched.
 93 Both varieties were represented by five replicates in each treatment. Detailed description of the experimental
 set-up is given in Hansen et al. (2019), Frenck et al. (2011) and Ingvordsen et al. (2015).

Treatment	Temperature, day/night ($^{\circ}\text{C}$)	Humidity, day/night (%)	[O ₃] (ppb)	[CO ₂] (ppm)
Control	$19.4 \pm 2.5 / 13.8 \pm 4.1$	$53.7 \pm 5.3 / 65.8 \pm 8.3$	6.4 ± 2.1	534 ± 109
Episodic	$19.4 \pm 2.5 / 14.0 \pm 4.1$	$54.2 \pm 5.2 / 65.5 \pm 8.0$	84.5 ± 28.1	539 ± 109
Chronic	$19.4 \pm 2.5 / 13.9 \pm 4.1$	$53.7 \pm 5.4 / 65.5 \pm 8.3$	78.8 ± 32.4	537 ± 111

Table 1: Mean and standard deviation of growing conditions in RERAF chambers for wheat cultivars Lennox and KWS Bittern.

94

95 2.2. Ozone treatments

96 O₃ was generated by UV Pro 550 A ozone generators (Crystal Air Products and Services, Langley, BC,
 97 Canada). The experiments included 3 levels of fumigation: i) no O₃ enrichment (Control); ii) episodic O₃
 98 exposure (Episodic) ; and iii) full-time O₃ exposure (Chronic) (Fig. 1 and Table 1). In the Control treatment,
 99 O₃ concentration in the climate chambers was 6.4 ± 2.1 ppb during the whole crop cycle. In the Chronic
 100 treatment, the plants were exposed to 78.8 ± 32.4 ppb O₃ concentration during the daylight hours from
 101 sowing (Zadoks Stage 01 - ZS 01) to harvest maturity (ZS 99). In the Episodic treatment, O₃ concentration
 102 was 84.5 ± 28.1 ppb during the daylight hours and the duration of plant exposure was from the first node stage
 103 (ZS 31) until anthesis was complete (ZS 69). During the night, in both the Chronic and Episodic treatment,
 104 O₃ concentration was reduced to the Control level.

105 2.3. Plant measurements and calculation of evapotranspiration and water use efficiency

106 At the end of the experiment, the plants were harvested and dried for 48 h at 80°C . The above-ground
 107 biomass and grain yield were measured in g pot^{-1} and were converted to g m^{-2} using the pot dimensions. This
 108 allowed direct comparison between the observations and the model output. The plant water consumption (g
 109 pot^{-1}) was calculated as the difference in pot weight between two consecutive measurements. Assuming that
 110 the increase in plant weight between two measurements was negligible, we calculated canopy evapotranspi-
 111 ration (ET) (mm) as following:

$$ET = ((W_{p_n} - W_{p_{n+1}}) / \rho \cdot \text{pot}) \cdot 1000 \text{ mm/m} \quad (1)$$

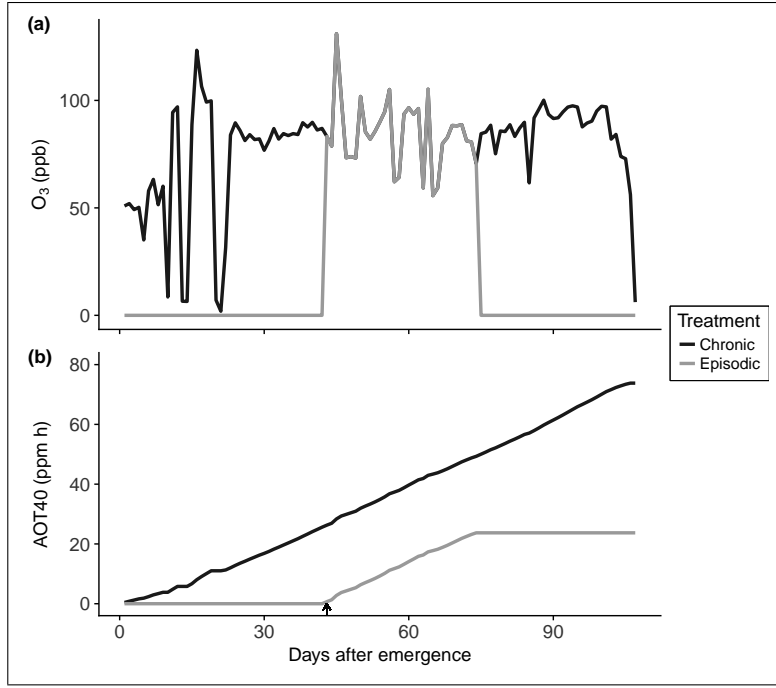


Figure 1: (a) Daily mean O₃ concentration (ppb) and (b) cumulative O₃ exposure above 40 ppb (AOT40) calculated from hourly [O₃] in Chronic and Episodic treatment. Arrow indicates day when plants reached ZS 31 (Zadoks stage 31).

112 where W_{p_n} and $W_{p_{n+1}}$ are the pot weight directly after the n irrigation (kg) and directly before the $n+1$ irrigation
 113 (kg) respectively, p is the number of pots per m² and ρ is water density (997 kg m³). Harvest index (HI) was
 114 calculated as the ratio of grain yield to above-ground biomass.

115 The biomass, grain yield, ET, HI and water use efficiency (WUE) of Lennox and KWS Bittern wheat in the
 116 experiments were calculated as the mean of the 5 replicates. The replicate 2 of Lennox in the Control and the
 117 replicate 5 of KWS Bittern in the Chronic treatment were disregarded due to errors in the measurements of
 118 pot weight. Also WUE (g m⁻² mm⁻¹) was defined as the ratio of the above-ground biomass (g m⁻²) to total ET
 119 (mm) at harvest.

120 *2.4. Ozone metrics*

121 The AOT40 index (Accumulated ozone exposure above a threshold of 40 ppbv) was calculated as follows:

$$AOT40 = \sum_{i=1}^n DOE40_i \tag{2}$$

122 where n is the number of days in the growing season, i is the day index and DOE40 is the daily O₃ exposure
 123 (ppm h) defined as:

$$DOE40 = \sum_{j=1}^m \max([O_3]_j - 40ppb, 0) \tag{3}$$

124 where [O₃] is the one hour mean O₃ concentration (ppb), m is the number of daylight hours per day and j is
 125 the hour index.

126 *2.5. GLAM-Parti model*

127 The GLAM-Parti crop model was developed based on the General Large Area Model for annual crops
128 (GLAM) which is a relatively simple model designed to operate at regional scale (Challinor et al., 2004). The
129 model was selected for the incorporation of the effect of O₃ on wheat, since it was developed with the SEMAC
130 approach (Simultaneous Equation Modelling for Annual Crops), a novel crop modelling methodology which
131 provides with a consistent representation of abiotic stresses and ensures internal consistency in the simulation
132 of crop growth and development under environmental stress conditions (Droutsas et al., 2019). GLAM-Parti
133 uses transpiration efficiency to simulate crop growth and allometric relationships for partitioning the biomass to
134 the plant compartments. The daily potential evapotranspiration is calculated by the Priestley-Taylor approach
135 and is partitioned into potential evaporation and potential transpiration. The actual transpiration is computed
136 from the potential transpiration rate by taking into account the soil water content. The transpiration is multiplied
137 by the transpiration efficiency to return the daily biomass growth.

138 Two major modifications were implemented into GLAM-Parti for this study. Firstly, the canopy SLA was
139 expressed as function of LAI (see Appendix A.1). In addition, the plant biomass partitioning scheme with
140 allometric relationships was extended to the post-anthesis period (see Appendix A.2). This method replaced
141 the previously used GLAM approach for simulating crop growth and development after anthesis in GLAM-Parti.

142 *2.6. GLAM-ROC development*

143 GLAM-ROC is the version of GLAM-Parti which incorporates the effect of O₃ on crop growth and devel-
144 opment. The O₃ damage to wheat was introduced into the model by reducing the canopy ET as well as
145 transpiration efficiency (TE) in daily time step. The effect of O₃ on ET was related to the AOT40 index and
146 the reduction in TE was expressed as function of [O₃]. An acclimation factor was introduced to simulate the
147 plant adjustment to stress conditions with increased O₃ exposure. Harvest Index was also reduced to account
148 for decreased allocation of assimilates to the grains under exposure to enhanced O₃ during the grain-filling
149 period.

150 *2.6.1. Modelling ozone effects on evapotranspiration*

151 Plant exposure to O₃ decreases leaf transpiration due to stomatal closure (Temple, 1986; Bernacchi et al.,
152 2011), which may have widespread implications for atmospheric moisture and climate (Arnold et al., 2018;
153 Lombardozzi et al., 2012). Data analysis was conducted to examine the effect of O₃ on cumulative evapotran-

154 spiration (CET) during the exposure to the pollutant. CET was calculated as:

$$CET = \sum_{i=1}^n ET_i \quad (4)$$

155 where ET is canopy evapotranspiration, i is the day index and n is the number of days after planting. Data from
156 the variety Lennox was used to compare the differences in CET between the Control, Chronic and Episodic
157 treatment.

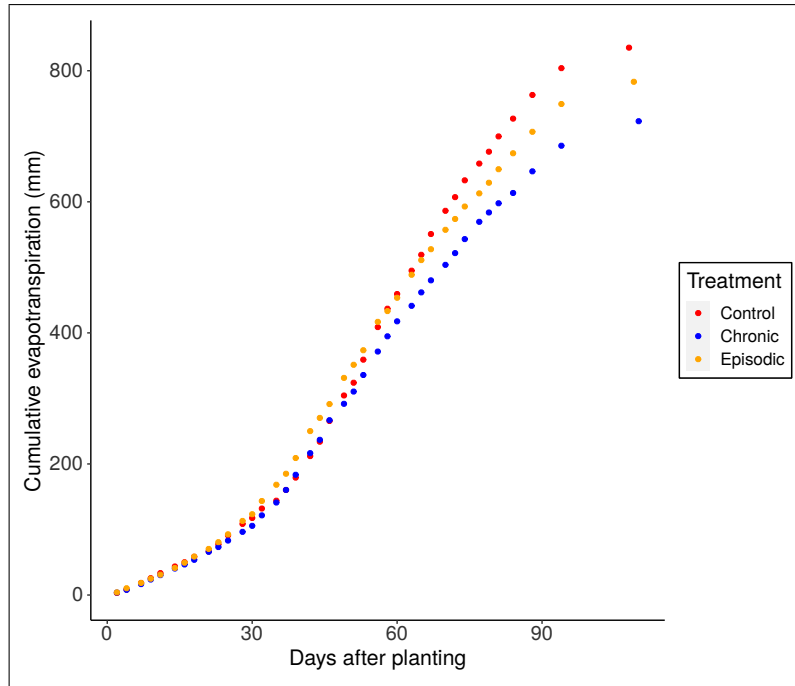


Figure 2: Cumulative evapotranspiration (CET) (mm) of wheat variety Lennox from planting to harvest for Control, Chronic and Episodic O₃ treatment.

158 CET exhibited a significant response to O₃ in both the Episodic and Chronic treatment, which showed 6.2%
159 and 13.4% lower end-of-season CET respectively in comparison with the Control (Fig. 2). Nevertheless, the
160 O₃ impact varied in magnitude with time and the plant sensitivity to O₃ was investigated according to the
161 growth stage. The crop cycle was separated into three stages, from seed germination (ZS 01) to the first
162 node stage (ZS 31) (Stage 1), the first node stage to the end of anthesis (ZS 69) (Stage 2) and from the end
163 of anthesis to harvest maturity (ZS 99) (Stage 3). We used the Pearson test to examine the differences in
164 CET between the Control and Chronic as well as Control and Episodic treatment (Table 2). During Stage 1,
165 there was a weak, non-significant correlation between the two variables (p-value >0.05). At that stage, only
166 the plants in the Chronic treatment were fumigated with O₃. This shows that the effect of O₃ before ZS 31
167 was not significant. On the other hand, during Stages 2 and 3 there was a significant positive correlation in
168 the difference in CET between Control and Chronic and Control and Episodic treatment (i.e. p-value < 0.001

169 in both stages). This implies that the O₃ impact was significant during Stages 2 and 3. Thus, the negative
 170 effects of O₃ on wheat were considered to initiate at the onset of stem elongation (ZS 31) until crop maturity.

	corr	p-value	Test
Stage 1	0.28	0.28	Pearson
Stage 2	0.98	<0.001	Pearson
Stage 3	0.97	<0.001	Pearson

Table 2: Correlation coefficients for difference in cumulative evapotranspiration (CET) between Control and Chronic as well as Control and Episodic O₃ treatment. Stage 1 is from seed germination (ZS 01) to first node (ZS 31), Stage 2 is from first node to end of anthesis (ZS 69) and Stage 3 is from end of anthesis to harvest maturity (ZS 99).

171 Next, we calculated the percentage change in CET (pCET) between the control and O₃-fumigated plants
 172 as follows:

$$173 \quad pCET_{oz} = (CET_{cc} - CET_{oz})/CET_{oz} \quad (5)$$

$$pCET_{ep.oz} = (CET_{cc} - CET_{ep.oz})/CET_{ep.oz} \quad (6)$$

174 where pCET_{oz} is the percentage change in CET between the Control and Chronic treatment and pCET_{ep.oz} is
 175 the percentage change in CET between the Control and Episodic treatment. Since only the differences after
 176 Stage 1 were considered, we normalized pCET_{oz} and pCET_{ep.oz} by subtracting their value at the end of Stage
 177 1. We also calculated the AOT40 index for the same time period (i.e. for Stages 2 and 3).

178 Fig. 3 (a) shows that the plants in the Episodic treatment were significantly more affected by the O₃
 179 exposure than the plants in the Chronic treatment and exhibited higher values of pCET. In other words, the
 180 plants which started in the low O₃ environment and were transferred to high O₃ at ZS 31 were more sensitive
 181 to the pollutant than the plants which grew at high [O₃] from emergence. Thus, the early fumigation with O₃ in
 182 the Chronic treatment decreased the plant sensitivity later in the season. This is in accordance with previous
 183 studies which report that the priming of plants can lead to improved performance at a subsequent abiotic
 184 stress event (Tanou et al., 2012; Wang et al., 2014; Li et al., 2014). The plants in the Episodic treatment were
 185 not fumigated with O₃ at Stage 1 and exhibited higher sensitivity to the pollutant during Stage 2.

186 2.6.2. Acclimation factor

187 The duration of plant exposure to O₃ affected the relationship between pCET and AOT40 (Fig. 3 (a)). We
 188 introduced the effective AOT40 index (efAOT40) which accounts for the variability in the effect of O₃ on wheat
 189 over time. The efAOT40 index represents the part of daily O₃ exposure which is limiting for the plant growth

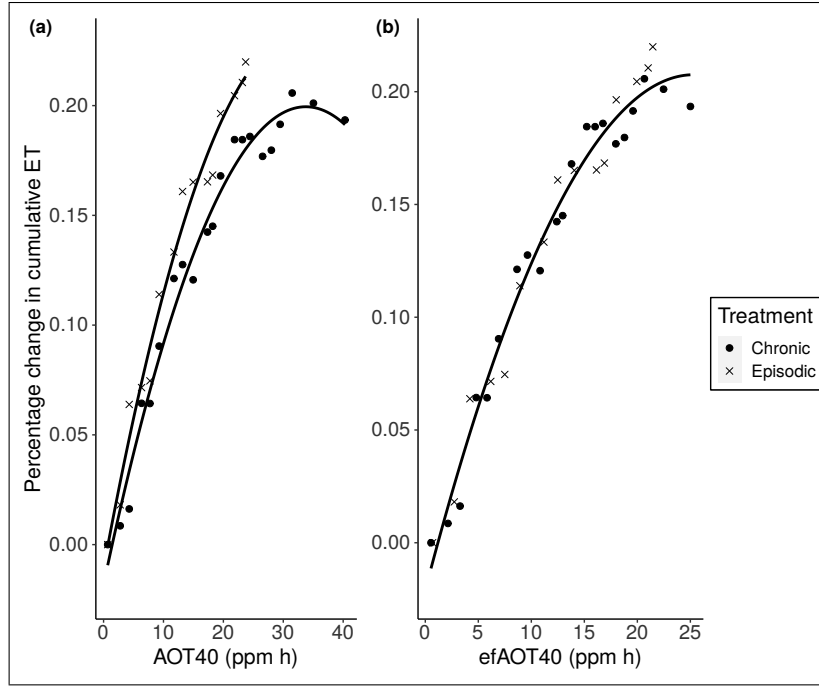


Figure 3: (a) Percentage change in cumulative evapotranspiration (pCET) of wheat variety Lennox between Control and Chronic treatment as well as Control and Episodic O₃ treatment plotted against AOT40; (b) pCET expressed as function of effective AOT40 (efAOT40) and continuous black line is the regression: $y = -0.021 + 0.018x - 0.000356x^2$ ($R^2=0.98$, $p < 0.01$). All pCET values were calculated for Stages 2, 3 after normalization at the end of Stage 1. AOT40 and efAOT40 were calculated for the same stages.

190 and is defined as:

$$efAOT40 = \sum_{i=1}^n (1 - f_{acl_i}) DOE40_i \quad (7)$$

191 where n is the number of days in the growing season, i is the day index and f_{acl} is an acclimation factor that
 192 accounts for the plant adjustment to O₃ over time. f_{acl} is a function of the number of days that DOE40 is above
 193 zero (ND_{oz}), it is in the [0,1] range and is updated in daily time step as follows:

$$f_{acl} = a_1 * f(ND_{oz}) \quad (8)$$

194 where a_1 is an empirical constant and ND_{oz} starts at zero at planting and is updated in daily time step as
 195 follows:

$$ND_{oz(i)} = \begin{cases} ND_{oz(i-1)} + 1 & DOE40_i > 0 \\ ND_{oz(i-1)} & DOE40_i = 0 \end{cases} \quad (9)$$

196 where i is the day after planting and $i-1$ is the previous day.

197 Due to incomplete understanding of the plant acclimation process to chronic O₃ stress, different equations
 198 were tested for the parameterization of f_{acl} . We evaluated the fit of a linear, quadratic and square root function
 199 in the expression of f_{acl} against ND_{oz} (Table 3). The parameter a_1 was calibrated to minimize RMSE for pCET

200 between the Chronic and Episodic treatment when expressed against efAOT40. RMSE was calculated as
 201 follows:

$$RMSE = \sqrt{\frac{\sum_{i=1}^n (pCET_{oz_i} - pCET_{ep,oz_i})^2}{n}} \quad (10)$$

202 where i is the day index and n is the number of observations.

203 For the derivation of Eq. 8, the linear shape was selected since it provided the lowest RMSE between
 204 all functions tested (Table 3). The relationship between pCET and efAOT40 was described by a second
 205 degree polynomial model (Fig. 3 (b)), which was used in GLAM-ROC to estimate the O₃-induced reduction
 206 on potential ET (i.e. the canopy ET rate under optimal growth conditions). Detailed information about the
 207 incorporation of the above-mentioned formula into the model is given in the Appendix A.3.

Line shape	Function	Calibrated value of a_1	RMSE
Linear	$a_1 ND_{oz}$	0.006	0.0124
Quadratic	$a_1 ND_{oz}^2$	0.0001	0.0182
Root	$a_1 \sqrt{ND_{oz}}$	0.05	0.0136

Table 3: Evaluation of different line shapes in the expression of acclimation factor (f_{acl}) as function of the number of days of O₃ exposure (ND_{oz}). The empirical parameter a_1 was calibrated to minimize RMSE between $pCET_{oz}$ and $pCET_{ep,oz}$ when expressed against efAOT40.

208 2.6.3. Ozone effects on transpiration efficiency and partitioning to grains

209 Exposure to O₃ induces up-regulation of the plant antioxidant metabolism which is energy demanding and
 210 the plants suppress their growth to use their resources for reducing the stress damage (Betzberger et al.,
 211 2010; Fatima et al., 2019). As a result TE decreases, since the plant growth reduction exceeds the reduction
 212 in ET (VanLoocke et al., 2012). HI also decreases due to reduced allocation of assimilates to the grains
 213 (Pleijel et al., 2014).

214 In GLAM-ROC, we applied O₃-induced modifications on both TE and the rate of increase of HI (dHI/dt).
 215 TE is defined as:

$$TE = \min\left(\frac{E_T}{VPD}, E_{TN,max}\right) \quad (11)$$

216 where E_T is normalised transpiration efficiency in Pa, VPD is vapour pressure deficit (kPa), and $E_{TN,max}$ is
 217 the maximum transpiration efficiency in $g\ kg^{-1}$. In this study, temperature and humidity were controlled (see
 218 Table 1) and VPD did not fluctuate significantly for most of the days in the growing season, thus for simplicity
 219 TE was set equal to $E_{TN,max}$. The effect of O₃ on TE was related to the effective [O₃] index (ef[O₃]). This
 220 index is calculated similarly to efAOT40 to simulate the plant adjustment to chronic O₃ stress which leads to

221 optimization of biomass productivity over time. $ef[O_3]$ is a fraction of daily $[O_3]$ defined as follows:

$$ef[O_3] = (1 - f_{acl}) \cdot [O_3] \quad (12)$$

222 where $[O_3]$ is the daily mean O_3 concentration during the daylight hours and f_{acl} is calculated in Eq. 8. For
 223 dHI/dt , no acclimation mechanism was assumed to impact on the allocation of assimilates to the grains, thus
 224 the effect was related to $[O_3]$.

225 The effects of O_3 on both TE and dHI/dt were initiated above 10 ppb which is the O_3 level of the pre-
 226 industrial period (Marenco et al., 1994). This threshold was set since GLAM-ROC is designed to simulate the
 227 effect of O_3 pollution on wheat in comparison with the pre-industrial period. TE and dHI/dt decreased linearly
 228 above the 10 ppb $[O_3]$ threshold as follows (Fig. 4): $ef[O_3]$ is a fraction of daily $[O_3]$ defined as follows:

$$\frac{TE_{oz}}{TE_c} = \begin{cases} 1 & [O_3] < 10ppb \\ c_1 \cdot ef[O_3] + d_1 & [O_3] \geq 10ppb \end{cases} \quad (13)$$

$$\frac{(dHI/dt)_{oz}}{(dHI/dt)_c} = \begin{cases} 1 & ef[O_3] < 10ppb \\ c_2 \cdot ef[O_3] + d_2 & ef[O_3] \geq 10ppb \end{cases} \quad (14)$$

229 where TE_c , TE_{oz} , $(dHI/dt)_c$ and $(dHI/dt)_{oz}$ are the control and O_3 -limited TE and dHI/dt respectively.

230 Feng et al. (2008) summarizes various experiments with wheat plants fumigated with different O_3 levels.
 231 The study finds that the aboveground biomass is decreased by an average of 18% at $[O_3]$ of 72 ppb in
 232 comparison with carbon-filtered treatments. Similarly, HI reduces by 9% at the same $[O_3]$ level. Following the
 233 above findings, the slope and intercept of Eq. 13, 14 were calculated accordingly and their values are given
 234 in Table A.5.

235 2.7. Model calibration and evaluation measures

236 The GLAM-ROC model was calibrated against the observed data for KWS Bittern wheat in the Control
 237 treatment. The metric used for the model calibration was the absolute error (AE) according to the following
 238 formula:

$$AE = |O - S| \quad (15)$$

239 where O and S are the observed and simulated values of the compared variables. The model performance
 240 was evaluated with the absolute percent error between the observed and simulated value of all compared
 241 variables.

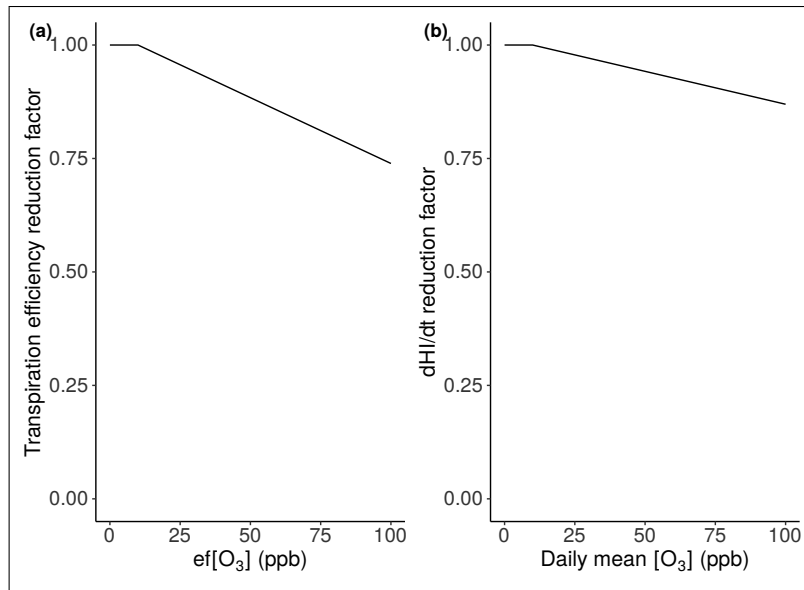


Figure 4: O₃-induced reduction (a) in transpiration efficiency (TE) relative to control expressed against effective daily mean [O₃] (ef[O₃]), (b) in the rate of increase of HI (dHI/dt) relative to control expressed as function of daily mean [O₃].

242 2.8. Calibration

243 The phenology of the model was set to meet the observed anthesis and maturity dates of the Control
 244 treatment. This was done to avoid any model bias from sources different than the O₃ stress effects. The
 245 maximum transpiration efficiency ($E_{TN,max}$) was calibrated with the use of an optimizer which selected the
 246 value that minimized AE between the end-of-season observed and simulated above-ground biomass in the
 247 Control treatment. Similarly, the rate of change of harvest index (dHI/dt) was selected by the optimizer to
 248 minimize AE between the observed and simulated grain yield of the Control treatment. The step for the runs
 249 of the optimizer was 0.1 for $E_{TN,max}$ and 0.0005 for dHI/dt. The ranges and values of the calibrated parameters
 250 are provided in Table A.6. All other parameter values were taken from Droutsas et al. (2019). The yield gap
 251 parameter (YGP) was set to one since O₃ was the only yield-limiting factor.

252 2.9. Sensitivity analysis

253 We performed a sensitivity analysis to test GLAM-ROC in a wide range of O₃ concentrations. The relative
 254 O₃ damage to yield was examined in comparison with the yield at the baseline O₃ concentration. In the meta-
 255 analysis of Pleijel et al. (2018), average [O₃] of charcoal-filtered air treatments was 13.3 ppb. We used the
 256 same baseline for direct comparison between the two studies. We modified all hourly O₃ concentrations in
 257 the Chronic and Episodic treatment by the appropriate value, such that the average [O₃] during the growing
 258 season was [13.3, 23.3, ..., 83.3 ppb]. We run GLAM-ROC at the different O₃ concentration levels by keeping

259 all other environmental conditions constant. The relative O₃ damages to yield in the Chronic and Episodic
260 treatment were calculated as percentage differences in yield from the baseline simulation.

261 **3. Results**

262 *3.1. Evaluation of GLAM-ROC model skill*

263 Exposure to O₃ significantly decreased the plant biomass and yield of KWS Bittern wheat in the exper-
264 iments as well as the total evapotranspiration (TET) and WUE. All measured and simulated values of the
265 compared variables and their percent error are shown in Table 4. The model reproduced the observed plant
266 biomass response in both O₃ treatments (Fig. 5(a)). In the Chronic treatment, the percent error between the
267 observed and simulated biomass at harvest was 5.02%. In the Episodic treatment, biomass was simulated to
268 within 1% of observation. Thus, the model closely followed the effect of O₃ stress on wheat biomass in both
269 durations of exposure by accounting for O₃-induced reductions in canopy ET and TE.

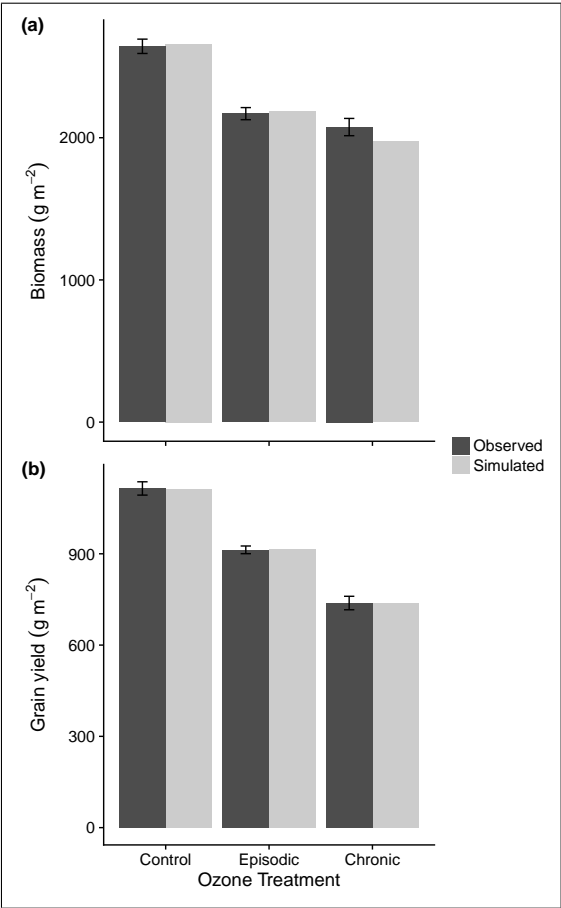


Figure 5: Observed (wheat variety KWS Bittern) and simulated (a) above-ground biomass and (b) grain yield at harvest in Control, Chronic and Episodic O₃ treatment. Error bars are standard errors of means in the observations.

270 Regarding the grain yield, GLAM-ROC accurately estimated the observed decreases in both the Chronic

	Control			Chronic			Episodic		
	Measured	Simulated	Percent error (%)	Measured	Simulated	Percent error (%)	Measured	Simulated	Percent error (%)
Biomass	2643.1	2659.0	0.60	2075.1	1971.0	5.02	2169.5	2185.0	0.71
Yield	1114.8	1113.0	0.16	738.2	738.0	0.03	913.0	915.0	0.22
HI	0.422	0.419	0.71	0.356	0.374	5.06	0.420	0.419	0.24
TET difference									
from control (%)	-	-	-	-12.10	-14.54	20.17	-9.42	-10.14	7.64
WUE difference									
from control (%)	-	-	-	-9.98	-13.27	32.97	-9.27	-8.55	7.77

Table 4: Measured and simulated above-ground biomass (g m^{-2}), grain yield (g m^{-2}), harvest index (HI), total ET (TET) and WUE difference from control (%) as well as their percent error in Control, Chronic and Episodic O_3 treatment.

271 and Episodic treatment. Yield was simulated to within 1% of observation in both the Chronic and Episodic
 272 treatment. Reduction in HI was also noticed in the Chronic but not the Episodic treatment (Fig. 6). This was
 273 due to lack of O_3 fumigation during the grain filling period in the Episodic treatment. The model reproduced
 274 the observed plant response to HI in the Episodic treatment where the simulated value was within 1% of
 275 observation. In the Chronic treatment, HI was overestimated by 5.06%.

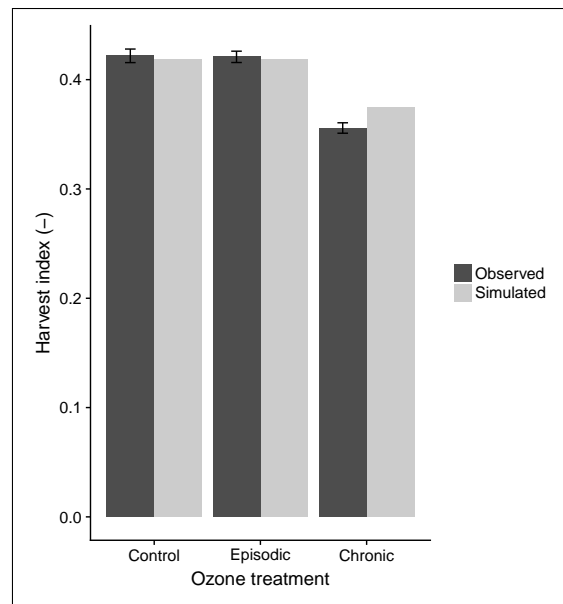


Figure 6: Observed (wheat variety KWS Bittern) and simulated harvest index in Control, Chronic and Episodic O_3 treatment. Error bars are standard errors of means in the observations.

276 Finally, GLAM-ROC overestimated the O_3 -induced reduction in TET in the Chronic treatment and the
 277 percent error was 20.17% (Fig. 7 (a)). The model skill was higher in the Episodic treatment where the percent
 278 error was 7.64%. WUE was significantly overestimated in the Chronic treatment, where the percent error was
 279 32.97 % (Fig. 7 (b)). In the Episodic treatment, GLAM-ROC exhibited improved skill and the percent error for

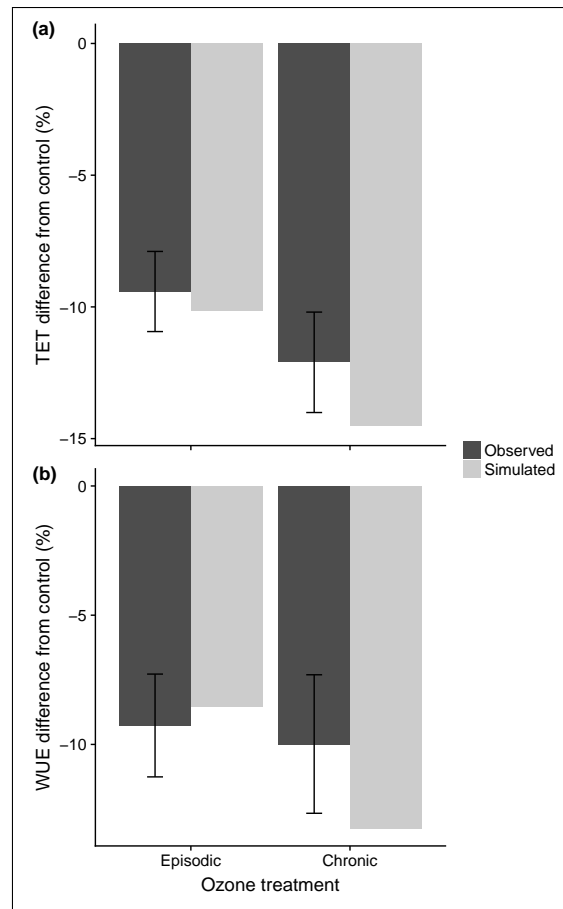


Figure 7: Observed (wheat variety KWS Bittern) and simulated difference from control in end-of season (a) total evapotranspiration (TET) and (b) water use efficiency (WUE) in Chronic and Episodic O₃ treatment. Error bars are standard errors of means in the observations.

281 **3.2. Sensitivity of GLAM-ROC to different O₃ concentrations**

282 Yield reduction was higher in the Chronic than the Episodic exposure in all O₃ concentrations (Fig. 8).
 283 In the Episodic treatment, yield reduced in an almost linear fashion for every 10 ppb increase in [O₃] from
 284 the baseline. Using linear regression of the data points, yield loss was found to increase by 0.28% per ppb
 285 increase in [O₃] (regression not shown). In the Chronic treatment, yield loss increased by 0.54% per ppb
 286 increase in [O₃] (regression not shown), however the standard error (se) of the slope was 177.3% higher than
 287 the Episodic treatment (i.e. se of slope was 0.0366 in Chronic against 0.0132 in Episodic treatment). This
 288 means that in the Chronic treatment, the reduction in grain yield diverted substantially from the linear line
 289 depending on [O₃]. The highest yield loss was estimated when the difference from the baseline [O₃] value
 290 increased from 30 to 40 ppb. In absolute numbers, the grain yield of wheat was most affected when [O₃]
 291 increased from 43.3 to 53.3 ppb. Within that concentration range, the average damage to yield was 0.96%
 292 per ppb increase in [O₃]. In the Episodic treatment, the same [O₃] range gave the highest damage to yield

293 with 0.39% loss per ppb increase in [O₃].

294 We also applied linear regression to all data points in the Chronic and Episodic treatment and compared
295 the regression line to those developed in the meta-analysis of Pleijel et al. (2018) and Broberg et al. (2015)
296 (Fig. 8). The three lines were in close agreement with each other and the slope of this study was -0.41 against
297 -0.36 of Pleijel et al. (2018) and -0.47 of Broberg et al. (2015). Thus, the studies suggest 0.41%, 0.36% and
298 0.47% increase in wheat yield loss respectively per ppb increase in [O₃] above the baseline. However, it
299 should be noted that the lower O₃ damage suggested by Pleijel et al. (2018) in comparison to Broberg et al.
300 (2015) may be explained by that the former study used wheat yield data only from charcoal-filtered and non-
301 filtered air treatments, whilst the latter study used also treatments with elevated O₃ levels. In addition, the
302 meta-analysis of Feng et al. (2008) estimates that the grain yield of wheat is reduced by 17.5 and 29% at
303 average [O₃] of 43 and 72 ppb respectively. Both findings are in very close agreement with the regression line
304 of our study which predicted the loss in grain yield with less than 3% difference from the reported values (Fig.
305 8). Overall, GLAM-ROC was in accordance with the existing meta-analysis studies and closely followed the
306 measured effect of O₃ on the grain yield of wheat.

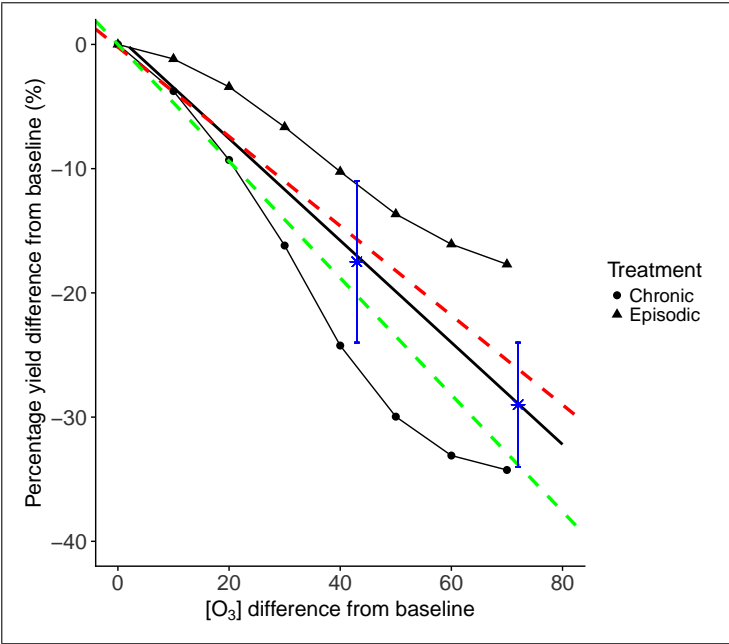


Figure 8: GLAM-ROC estimations of O₃-induced grain yield loss of wheat at different O₃ concentrations in Chronic (circles) and Episodic treatment (triangles) in comparison with baseline. Solid black line is linear regression of all data points in Chronic and Episodic treatment, red and green dashed lines are linear regressions in meta-analysis of Pleijel et al. (2018) and Broberg et al. (2015) respectively. Star data points are O₃-induced yield losses at 43 and 72 ppb [O₃] in meta-analysis of Feng et al. (2008) and error bars are 95% confidence intervals. The two star data points are presented using their absolute [O₃] value on x axis instead of the difference from baseline, since this was not reported.

307 3.3. GLAM-ROC comparison with ozone exposure response function

308 A large number of studies estimate the regional or global effect of O₃ on crop productivity based on a
309 statistical response function (SRF) between the relative crop yield and the level of O₃ exposure (e.g. Hollaway
310 et al., 2012; Ghude et al., 2014; Sharma et al., 2019). This formula assumes linear reduction in grain yield
311 in relation to AOT40. The modelling methodology introduced in GLAM-ROC assumes non-linear O₃ effect on
312 yield with increased exposure to the pollutant. The two methods were evaluated against the observed data for
313 KWS Bittern wheat. In the SRF model, the function for wheat was taken from Mills et al. (2007). The AOT40
314 index was accumulated during Stages 2 and 3, since these days were the most O₃-sensitive (i.e. the last 68
315 days of the crop season).

316 In GLAM-ROC, the grain yield in the Chronic treatment was 80.7% of the Episodic, which was less than
317 1% different from the observed value (Fig. 9). In the SRF model, the Chronic: Episodic yield ratio was 0.35,
318 underestimated by 56.5%. This was due to the overestimation of the O₃ damage to yield at a greater extent in
319 the Chronic than the Episodic treatment. Nevertheless, no acclimation mechanism is considered in the SRF
320 model and the O₃ damage to yield is linearly extrapolated as AOT40 increases. As a result, the observed
321 non-linear plant response with increased exposure to O₃ stress affected the skill of the model. Thus, GLAM-
322 ROC improved upon the SRF model in the estimation of the Chronic: Episodic yield ratio by accounting for
323 the plant acclimation to chronic O₃ stress at variable duration of exposure.

324 4. Discussion

325 We developed and evaluated the GLAM-ROC model to simulate the effect of O₃ on wheat growth and
326 development. The model successfully reproduced the O₃-induced damage to wheat biomass and yield in both
327 the Episodic and Chronic treatment. The plant biomass was simulated to within 6% of observation in both
328 durations of exposure. Similarly, the simulated grain yield was less than 1% different from the measurements.
329 The model also closely followed the observed effects of O₃ on HI, ET and WUE.

330 The modelling approach followed here is simpler than stomatal flux-based methods commonly used in crop
331 models. Such method was avoided since it strongly depends on stomatal conductance, a trait which is highly
332 complex (Buckley, 2017) and not simple to incorporate in process-based crop models. Difficulties may also be
333 faced in crop models with complex O₃ schemes regarding their parameterization for large scale applications
334 (Emberson et al., 2018). GLAM is a large area crop model and unwarranted complexity should be avoided
335 (Challinor et al., 2018). In addition, our approach – even relying on an exposure-based methodology –

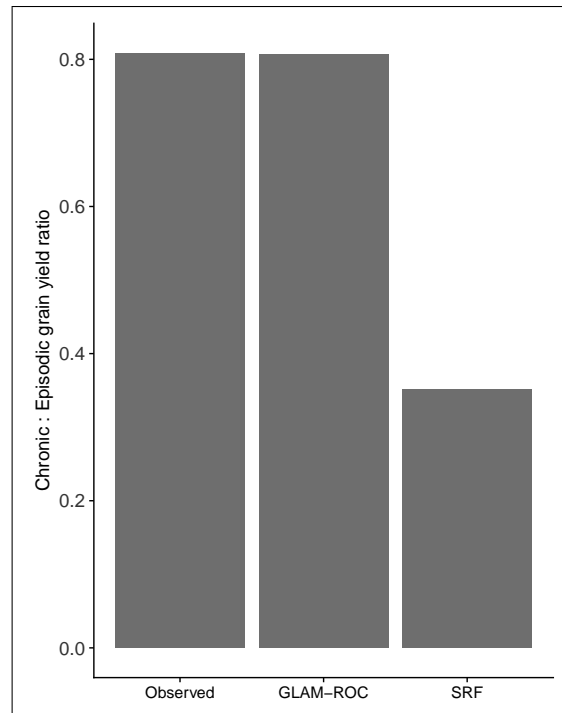


Figure 9: Observed and simulated grain yield in Chronic relative to Episodic treatment. SRF is statistical response function for wheat taken from Mills et al. (2007).

336 overcomes some limitations of the statistical response function. This is due to relating AOT40 to the potential
 337 ET rate instead of using the index to estimate grain yield loss directly. ET and AOT40 have been previously
 338 seen to correlate significantly under well-watered conditions (Jaudé et al., 2008; VanLoocke et al., 2012).
 339 Nevertheless, the use of AOT40 disregards the effect of O_3 stress on ET below 40 ppb. Bernacchi et al.
 340 (2011) and VanLoocke et al. (2012) exposed soybean plants to various O_3 concentrations and showed that
 341 the pollutant reduces canopy ET significantly for exposures above 40 ppb. Since wheat and soybean exhibit
 342 similar sensitivity to O_3 (Mills et al., 2007), in our study we also related the reduction in canopy ET to the
 343 AOT40 index.

344 In GLAM-ROC, the O_3 -induced decrease in ET is estimated in comparison with the same plant growing in
 345 optimal environment. Under water stress, the effect of O_3 on crop growth can be reduced due to decreased
 346 stomatal conductance and lower uptake of the pollutant by the leaves (Khan and Soja, 2003; Feng et al.,
 347 2008). Both O_3 and drought reduce the daily canopy transpiration rate in the model, the minimum of which
 348 is considered as the actual transpiration (i.e. the effect of drought (Challinor et al., 2004) and O_3 (this study)
 349 on ET are estimated independently). Thus, if limited soil water suppresses transpiration to a greater extend
 350 than O_3 , there will be no additive effect of the pollutant on canopy ET. In other words, the O_3 damage to crop
 351 transpiration and growth decreases with higher levels of water stress in GLAM-ROC. The accuracy of this

352 approach should be tested against experimental data with wheat exposed to both stressors simultaneously.
353 In this study, GLAM-ROC was only evaluated for the effect of O₃ on well-watered wheat crops. Thus, the
354 model can be currently used only in regions where adequate rainfall prevents water stress or where wheat
355 is fully irrigated. Elevated CO₂ concentrations can also reduce stomatal conductance and protect against
356 O₃ pollution (Yadav et al., 2019). Currently, GLAM-ROC does not account for the effect of elevated CO₂ on
357 crop growth and yield, thus it cannot be used for future environments with rising CO₂ concentrations (i.e. the
358 model has to be calibrated each time for the given CO₂ level). Following the addition of the CO₂ fertilization
359 mechanism, the interaction between elevated CO₂ and O₃ should be addressed to allow for the estimation of
360 crop performance and yield under future climate change conditions.

361 GLAM-ROC uses the acclimation factor to simulate the plant adaptation to chronic O₃ stress. This was
362 necessary for capturing the differences in water consumption and biomass productivity between the Chronic
363 and Episodic treatment. The plants in the Chronic treatment exhibited higher values of water consumption
364 than the Episodic during their common period of O₃ fumigation (i.e. at Stage 2). Nevertheless, only the plants
365 in Chronic treatment were exposed to high O₃ during Stage 1. The lack of previous exposure to the stressor in
366 the Episodic treatment increased the plant sensitivity at Stage 2, thus reducing the water consumption rates.
367 This difference in plant behaviour could not be simulated without considering the effect of plant acclimation to
368 chronic O₃ exposure. The acclimation factor was calculated according to the number of days of O₃ fumigation,
369 thus modifying the plant sensitivity to O₃ at different durations of exposure. The same factor simulated the
370 differences in biomass productivity between the two O₃ exposures through modifying the effect on TE. The
371 SRF model does not account for the plant acclimation mechanism and overestimated the relative damage to
372 grain yield between the Chronic and Episodic treatment by 56.5%.

373 In GLAM-ROC, the acclimation factor was related to the 40 ppb threshold (Eq. 8, 9), which means that
374 no plant acclimation was considered for exposure to O₃ below that level. This threshold was chosen since
375 it is the most commonly used threshold for relating the O₃ exposure to loss in crop yield (e.g., Fuhrer et al.,
376 1997; Mills et al., 2011; Sharma et al., 2019). However, it is unclear if this is the optimal threshold for wheat
377 acclimation to the pollutant or if it needs to be adjusted in the future. In our study, the O₃ concentrations in
378 the chambers were either very low (below 10 ppb in the Control treatment) or significantly higher than 40 ppb
379 during exposure to the pollutant in the Chronic and Episodic treatments. Thus, decreasing the acclimation
380 threshold does not change the model results in comparison with the observations (Section 3.1). In addition,
381 the sensitivity analysis (Section 3.2) indicates that this threshold is appropriate, since the model can be reliably

382 used for simulating grain yield losses for O₃ exposure below 40 ppb. Nevertheless, since wheat performance
383 is affected by O₃ below 40 ppb (Pleijel, 2011), the plant acclimation threshold may need to be reconsidered in
384 the future.

385 The plant sensitivity to O₃ varies also according to the growth stage. O₃ did not exhibit significant effect
386 on wheat from plant emergence to ZS 31, at least in terms of water consumption. Thus, the O₃ damage to
387 wheat was simulated to initiate at ZS 31. On the other hand, the period from anthesis to the end of grain filling
388 is the most O₃-sensitive for grain yield reduction (Lee et al., 1988; Pleijel et al., 1998; Soja et al., 2000). The
389 Episodic treatment was not fumigated with high O₃ after anthesis and HI was less than 1% different from the
390 Control. In the Chronic treatment, HI was 14.3% lower than the Control due to high O₃ exposure during grain
391 filling. GLAM-ROC was able to reproduce the O₃-induced reduction to yield by slowing down the daily rate of
392 increase of HI (dHI/dt) based on the [O₃] level. As a result, the model followed the observed decrease in HI
393 and successfully simulated the O₃ impact on grain yield at maturity.

394 The development and evaluation of GLAM-ROC were based on controlled-environment chamber exper-
395 iments where the environmental conditions cannot perfectly match the field. For instance, the plants were
396 grown in pots instead of being rooted on the ground. Nevertheless, the meta-analysis of Feng and Kobayashi
397 (2009) revealed no significant differences in the yield response to O₃ between pot and ground-rooted wheat
398 plants. In addition, the daily O₃ concentration in the chambers did not match the diurnal variation experi-
399 enced under ambient conditions (e.g. Pawlak and Jarosławski, 2015; Wang et al., 2017). However, Harmens
400 et al. (2018) exposed a modern wheat variety to various background O₃ concentrations and different peak O₃
401 episodes and showed that the relationship between the reduction in grain yield and the accumulated stom-
402 atal O₃ flux could be explained by the same slope irrespective of the temporal O₃ profile. Moreover, in our
403 study the fumigation in the Chronic treatment lasted for the full crop cycle (i.e. 107 days) with an average
404 concentration of 78.8 ppb, which can be unrealistic for most parts of the world. The highest frequency of O₃
405 pollution episodes (i.e. daily average 8-h [O₃] of at least 75 ppb) in the summertime for the year 2000 was 38
406 days in North America (Lei et al., 2012). The Episodic treatment was closer to these conditions with the total
407 duration of plant O₃ exposure being 33 days. The real dynamics of surface O₃ in polluted regions are likely
408 to be between the Episodic and Chronic treatments of this study. In addition, the average CO₂ concentration
409 in the chambers was around 530 ppm and this concentration level was experienced in all treatments. For this
410 reason, CO₂ was not accounted as an additional varying factor in the estimation of O₃ damage to wheat.

411 Our sensitivity analysis suggested 0.41% average yield loss per ppb increase in O₃ concentration above

412 13.3 ppb, which is directly comparable with existing meta-analysis studies (Fig. 8). In the Chronic treatment,
413 the model estimated a 0.96% maximum damage to yield per ppb increase in O₃ concentration, which occurred
414 when [O₃] was in the 43.3 - 53.3 ppb range. In the Episodic treatment, the same concentration range gave
415 the maximum damage to yield per ppb increase in [O₃], which was considerably lower at 0.39%. Overall, the
416 yield loss per ppb increase in [O₃] varied from 0.17% to 0.96%, depending on the treatment (i.e. Chronic vs.
417 Episodic) and the [O₃] level. Hence, the duration of exposure to O₃ stress is a significant factor influencing
418 the effect of the pollutant on wheat productivity and yield. Longer duration of exposure to O₃ implies higher
419 reduction in yield, however the relative damage may decrease as the duration increases. The non-linear
420 grain yield vs. [O₃] pattern can result from enhanced plant acclimation with increased duration exposure to
421 the pollutant. Thus, we believe that the plant acclimation process should be taken into account for robust
422 estimation of the chronic effect of O₃ on crop growth, productivity and yield.

423 **5. Conclusion**

424 Exposure to O₃ significantly decreased the wheat biomass and grain yield in the experiments. The GLAM-
425 ROC crop model was developed and evaluated for the effect of O₃ on wheat growth and development. A
426 statistical relationship was introduced to estimate the reduction in canopy evapotranspiration based on the O₃
427 exposure (i.e. AOT40 index). Decreases in transpiration efficiency and harvest index were also incorporated
428 into the model according to the O₃ concentration. The model successfully reproduced the observed O₃
429 damage to biomass and yield of KWS Bittern wheat in both the Episodic and Chronic treatment. Accounting
430 for the plant acclimation to chronic O₃ stress was necessary for good model skill. The acclimation process
431 was empirically incorporated with the use of an acclimation factor based on the days of O₃ exposure. This
432 allowed the simulation of plant adjustment to O₃ over time which reduced the relative damage to biomass
433 and yield. The statistical response function ignored the acclimation process and overestimated the Chronic:
434 Episodic grain yield ratio. It is concluded that the plant acclimation to chronic O₃ stress is significant and
435 should be taken into account for the estimation of the O₃ damage to wheat growth and productivity.

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442 **References**

443 Ainsworth, E. A., 2017. Understanding and improving global crop response to ozone pollution. *The Plant*
444 *Journal* 90 (5), 886–897.

445 Arnold, S., Lombardozzi, D., Lamarque, J.-F., Richardson, T., Emmons, L., Tilmes, S., Sitch, S., Folberth,
446 G., Hollaway, M., Val Martin, M., 2018. Simulated global climate response to tropospheric ozone-induced
447 changes in plant transpiration. *Geophysical Research Letters* 45 (23), 13–070.

448 Ashmore, M., 2005. Assessing the future global impacts of ozone on vegetation. *Plant, Cell & Environment*
449 28 (8), 949–964.

450 Avnery, S., Mauzerall, D. L., Liu, J., Horowitz, L. W., 2011. Global crop yield reductions due to surface ozone
451 exposure: 1. year 2000 crop production losses and economic damage. *Atmospheric Environment* 45 (13),
452 2284–2296.

453 Barnes, J., Velissariou, D., Davison, A., Holevas, C., 1990. Comparative ozone sensitivity of old and modern
454 greek cultivars of spring wheat. *New Phytologist* 116 (4), 707–714.

455 Bernacchi, C. J., Leakey, A. D., Kimball, B. A., Ort, D. R., 2011. Growth of soybean at future tropospheric
456 ozone concentrations decreases canopy evapotranspiration and soil water depletion. *Environmental pollu-*
457 *tion* 159 (6), 1464–1472.

458 Betzelberger, A. M., Gillespie, K. M., Mcgrath, J. M., Koester, R. P., Nelson, R. L., Ainsworth, E. A., 2010.
459 Effects of chronic elevated ozone concentration on antioxidant capacity, photosynthesis and seed yield of
460 10 soybean cultivars. *Plant, Cell & Environment* 33 (9), 1569–1581.

461 Borrell, A. K., Incoll, L., Simpson, R. J., Dalling, M. J., 1989. Partitioning of dry matter and the deposition and
462 use of stem reserves in a semi-dwarf wheat crop. *Annals of Botany* 63 (5), 527–539.

463 Broberg, M. C., Feng, Z., Xin, Y., Pleijel, H., 2015. Ozone effects on wheat grain quality—a summary. *Environ-*
464 *mental pollution* 197, 203–213.

465 Bruce, T. J., Matthes, M. C., Napier, J. A., Pickett, J. A., 2007. Stressful “memories” of plants: evidence and
466 possible mechanisms. *Plant Science* 173 (6), 603–608.

467 Buckley, T. N., 2017. Modeling stomatal conductance. *Plant physiology*, pp–01772.

468 Buckley, T. N., Mott, K. A., 2013. Modelling stomatal conductance in response to environmental factors. *Plant,*
469 *cell & environment* 36 (9), 1691–1699.

470 Burney, J., Ramanathan, V., 2014. Recent climate and air pollution impacts on indian agriculture. *Proceedings*
471 *of the National Academy of Sciences* 111 (46), 16319–16324.

472 Challinor, A., Wheeler, T., Craufurd, P., Slingo, J., Grimes, D., 2004. Design and optimisation of a large-area
473 process-based model for annual crops. *Agricultural and forest meteorology* 124 (1-2), 99–120.

474 Challinor, A. J., Ewert, F., Arnold, S., Simelton, E., Fraser, E., 2009. Crops and climate change: progress,
475 trends, and challenges in simulating impacts and informing adaptation. *Journal of experimental botany*
476 60 (10), 2775–2789.

477 Challinor, A. J., Müller, C., Asseng, S., Deva, C., Nicklin, K. J., Wallach, D., Vanuytrecht, E., Whitfield, S.,
478 Ramirez-Villegas, J., Koehler, A.-K., 2018. Improving the use of crop models for risk assessment and
479 climate change adaptation. *Agricultural systems* 159, 296–306.

480 Chen, C. P., Frei, M., Wissuwa, M., 2011. The *ozt8* locus in rice protects leaf carbon assimilation rate and
481 photosynthetic capacity under ozone stress. *Plant, cell & environment* 34 (7), 1141–1149.

482 Christy, B., Tausz-Posch, S., Tausz, M., Richards, R., Rebetzke, G., Condon, A., McLean, T., Fitzgerald, G.,
483 Bourgault, M., O'leary, G., 2018. Benefits of increasing transpiration efficiency in wheat under elevated CO_2
484 for rainfed regions. *Global change biology* 24 (5), 1965–1977.

485 Damour, G., Simonneau, T., Cochard, H., Urban, L., 2010. An overview of models of stomatal conductance at
486 the leaf level. *Plant, Cell & Environment* 33 (9), 1419–1438.

487 Droutsas, I., Challinor, A., Swiderski, M., Semenov, M. A., 2019. New modelling technique for improving crop
488 model performance-application to the glam model. *Environmental Modelling & Software* 118, 187–200.

489 Emberson, L., Ashmore, M., Cambridge, H., Simpson, D., Tuovinen, J.-P., 2000. Modelling stomatal ozone
490 flux across europe. *Environmental Pollution* 109 (3), 403–413.

491 Emberson, L., Büker, P., Ashmore, M., Mills, G., Jackson, L., Agrawal, M., Atikuzzaman, M., Cinderby, S.,
492 Engardt, M., Jamir, C., et al., 2009. A comparison of north american and asian exposure–response data for
493 ozone effects on crop yields. *Atmospheric Environment* 43 (12), 1945–1953.

494 Emberson, L. D., Pleijel, H., Ainsworth, E. A., Van den Berg, M., Ren, W., Osborne, S., Mills, G., Pandey, D.,
495 Dentener, F., B ker, P., et al., 2018. Ozone effects on crops and consideration in crop models. *European*
496 *Journal of Agronomy* 100, 19–34.

497 Ewert, F., Porter, J. R., 2000. Ozone effects on wheat in relation to co2: modelling short-term and long-term
498 responses of leaf photosynthesis and leaf duration. *Global Change Biology* 6 (7), 735–750.

499 FAO, I., UNICEF, et al., 2017. Wfp, who (2017) the state of food security and nutrition in the world 2017.
500 Building Resilience for Peace and Food Security (Food and Agriculture Organization, Rome).

501 Farage, P., Long, S. P., 1999. The effects of o3 fumigation during leaf development on photosynthesis of wheat
502 and pea: an in vivo analysis. *Photosynthesis Research* 59 (1), 1–7.

503 Farage, P. K., Long, S. P., Lechner, E. G., Baker, N. R., 1991. The sequence of change within the photosyn-
504 thetic apparatus of wheat following short-term exposure to ozone. *Plant Physiology* 95 (2), 529–535.

505 Fatima, A., Singh, A. A., Mukherjee, A., Agrawal, M., Agrawal, S. B., 2019. Ascorbic acid and thiols as
506 potential biomarkers of ozone tolerance in tropical wheat cultivars. *Ecotoxicology and environmental safety*
507 171, 701–708.

508 Feng, Z., Kobayashi, K., 2009. Assessing the impacts of current and future concentrations of surface ozone
509 on crop yield with meta-analysis. *Atmospheric Environment* 43 (8), 1510–1519.

510 Feng, Z., Kobayashi, K., Ainsworth, E. A., 2008. Impact of elevated ozone concentration on growth, physiology,
511 and yield of wheat (*triticum aestivum* L.): a meta-analysis. *Global Change Biology* 14 (11), 2696–2708.

512 Feng, Z., Wang, L., Pleijel, H., Zhu, J., Kobayashi, K., 2016. Differential effects of ozone on photosynthesis of
513 winter wheat among cultivars depend on antioxidative enzymes rather than stomatal conductance. *Science*
514 *of the Total Environment* 572, 404–411.

515 Fiscus, E. L., Booker, F. L., Burkey, K. O., 2005. Crop responses to ozone: uptake, modes of action, carbon
516 assimilation and partitioning. *Plant, Cell & Environment* 28 (8), 997–1011.

517 Frenck, G., van der Linden, L., Mikkelsen, T. N., Brix, H., J rgensen, R. B., 2011. Increased [co2] does
518 not compensate for negative effects on yield caused by higher temperature and [o3] in *brassica napus* L.
519 *European Journal of Agronomy* 35 (3), 127–134.

520 Fuhrer, J., Skärby, L., Ashmore, M. R., 1997. Critical levels for ozone effects on vegetation in Europe. *Environmental Pollution* 97 (1-2), 91–106.

521

522 Ghude, S. D., Jena, C., Chate, D., Beig, G., Pfister, G., Kumar, R., Ramanathan, V., 2014. Reductions in
523 India's crop yield due to ozone. *Geophysical Research Letters* 41 (15), 5685–5691.

524 Gillespie, K. M., Rogers, A., Ainsworth, E. A., 2011. Growth at elevated ozone or elevated carbon dioxide
525 concentration alters antioxidant capacity and response to acute oxidative stress in soybean (*Glycine max*).
526 *Journal of Experimental Botany* 62 (8), 2667–2678.

527 Gillespie, K. M., Xu, F., Richter, K. T., McGrath, J. M., Markelz, R. C., Ort, D. R., Leakey, A. D., Ainsworth,
528 E. A., 2012. Greater antioxidant and respiratory metabolism in field-grown soybean exposed to elevated O₃
529 under both ambient and elevated CO₂. *Plant, Cell & Environment* 35 (1), 169–184.

530 Hansen, E. M., Hauggaard-Nielsen, H., Launay, M., Rose, P., Mikkelsen, T. N., 2019. The impact of ozone
531 exposure, temperature and CO₂ on the growth and yield of three spring wheat varieties. *Environmental and*
532 *Experimental Botany*, 103868.

533 Harmens, H., Hayes, F., Mills, G., Sharps, K., Osborne, S., Pleijel, H., 2018. Wheat yield responses to
534 stomatal uptake of ozone: Peak vs rising background ozone conditions. *Atmospheric Environment* 173,
535 1–5.

536 Heath, R. L., 1994. Possible mechanisms for the inhibition of photosynthesis by ozone. *Photosynthesis Re-*
537 *search* 39 (3), 439–451.

538 Heath, R. L., Lefohn, A. S., Musselman, R. C., 2009. Temporal processes that contribute to nonlinearity in
539 vegetation responses to ozone exposure and dose. *Atmospheric Environment* 43 (18), 2919–2928.

540 Held, A., Mooney, H., Gorham, J. N., 1991. Acclimation to ozone stress in radish: leaf demography and
541 photosynthesis. *New Phytologist* 118 (3), 417–423.

542 Hollaway, M. J., Arnold, S., Challinor, A. J., Emberson, L., 2012. Intercontinental trans-boundary contributions
543 to ozone-induced crop yield losses in the northern hemisphere. *Biogeosciences* 9 (1), 271–292.

544 Ingvordsen, C. H., Backes, G., Lyngkjær, M. F., Peltonen-Sainio, P., Jensen, J. D., Jalli, M., Jahoor, A.,
545 Rasmussen, M., Mikkelsen, T. N., Stockmarr, A., et al., 2015. Significant decrease in yield under future
546 climate conditions: Stability and production of 138 spring barley accessions. *European Journal of Agronomy*
547 63, 105–113.

548 Jaudé, M. B., Katerji, N., Mastrorilli, M., Rana, G., 2008. Analysis of the effect of ozone on soybean in the
549 mediterranean region: I. the consequences on crop-water status. *European Journal of Agronomy* 28 (4),
550 508–518.

551 Khan, S., Soja, G., 2003. Yield responses of wheat to ozone exposure as modified by drought-induced differ-
552 ences in ozone uptake. *Water, Air, and Soil Pollution* 147 (1-4), 299–315.

553 Kollist, H., Zandalinas, S. I., Sengupta, S., Nuhkat, M., Kangasjärvi, J., Mittler, R., 2018. Rapid responses to
554 abiotic stress: priming the landscape for the signal transduction network. *Trends in plant science*.

555 Lee, E. H., Tingey, D. T., Hogsett, W. E., 1988. Evaluation of ozone exposure indices in exposure-response
556 modeling. *Environmental Pollution* 53 (1-4), 43–62.

557 Lei, H., Wuebbles, D. J., Liang, X.-Z., 2012. Projected risk of high ozone episodes in 2050. *Atmospheric*
558 *environment* 59, 567–577.

559 Li, X., Cai, J., Liu, F., Dai, T., Cao, W., Jiang, D., 2014. Cold priming drives the sub-cellular antioxidant
560 systems to protect photosynthetic electron transport against subsequent low temperature stress in winter
561 wheat. *Plant Physiology and Biochemistry* 82, 34–43.

562 Lombardozi, D., Levis, S., Bonan, G., Sparks, J., 2012. Predicting photosynthesis and transpiration re-
563 sponses to ozone: decoupling modeled photosynthesis and stomatal conductance. *Biogeosciences Dis-*
564 *cussions* 9 (4).

565 Marenco, A., Gouget, H., Nédélec, P., Pagés, J.-P., Karcher, F., 1994. Evidence of a long-term increase
566 in tropospheric ozone from pic du midi data series: Consequences: Positive radiative forcing. *Journal of*
567 *Geophysical Research: Atmospheres* 99 (D8), 16617–16632.

568 Mauzerall, D. L., Wang, X., 2001. Protecting agricultural crops from the effects of tropospheric ozone expo-
569 sure: reconciling science and standard setting in the united states, europe, and asia. *Annual Review of*
570 *energy and the environment* 26 (1), 237–268.

571 McGrath, J. M., Betzelberger, A. M., Wang, S., Shook, E., Zhu, X.-G., Long, S. P., Ainsworth, E. A., 2015. An
572 analysis of ozone damage to historical maize and soybean yields in the united states. *Proceedings of the*
573 *National Academy of Sciences* 112 (46), 14390–14395.

574 Mikkelsen, T. N., Ro-Poulsen, H., 1994. Exposure of norway spruce to ozone increases the sensitivity of
575 current year needles to photoinhibition and desiccation. *New Phytologist* 128 (1), 153–163.

576 Mills, G., Buse, A., Gimeno, B., Bermejo, V., Holland, M., Emberson, L., Pleijel, H., 2007. A synthesis of aot40-
577 based response functions and critical levels of ozone for agricultural and horticultural crops. *Atmospheric*
578 *Environment* 41 (12), 2630–2643.

579 Mills, G., Hayes, F., Simpson, D., Emberson, L., Norris, D., Harmens, H., Büker, P., 2011. Evidence of
580 widespread effects of ozone on crops and (semi-) natural vegetation in europe (1990–2006) in relation
581 to aot40-and flux-based risk maps. *Global Change Biology* 17 (1), 592–613.

582 Mills, G., Sharps, K., Simpson, D., Pleijel, H., Frei, M., Burkey, K., Emberson, L., Uddling, J., Broberg, M.,
583 Feng, Z., et al., 2018. Closing the global ozone yield gap: Quantification and cobenefits for multistress
584 tolerance. *Global change biology* 24 (10), 4869–4893.

585 Moot, D., Jamieson, P., Henderson, A., Ford, M., Porter, J., 1996. Rate of change in harvest index during
586 grain-filling of wheat. *The Journal of Agricultural Science* 126 (4), 387–395.

587 Mulholland, B., Craighan, J., Black, C., Colls, J., Atherton, J., Landon, G., 1998. Effects of elevated co2 and
588 o3 on the rate and duration of grain growth and harvest index in spring wheat (*triticum aestivum* L.). *Global*
589 *Change Biology* 4 (6), 627–635.

590 Musselman, R. C., Lefohn, A. S., Massman, W. J., Heath, R. L., 2006. A critical review and analysis of the
591 use of exposure-and flux-based ozone indices for predicting vegetation effects. *Atmospheric Environment*
592 40 (10), 1869–1888.

593 Pawlak, I., Jarosławski, J., 2015. The influence of selected meteorological parameters on the concentration
594 of surface ozone in the central region of poland. *Atmosphere-Ocean* 53 (1), 126–139.

595 Pleijel, H., 2011. Reduced ozone by air filtration consistently improved grain yield in wheat. *Environmental*
596 *pollution* 159 (4), 897–902.

597 Pleijel, H., Broberg, M. C., Uddling, J., Mills, G., 2018. Current surface ozone concentrations significantly
598 decrease wheat growth, yield and quality. *Science of the Total Environment* 613, 687–692.

599 Pleijel, H., Danielsson, H., Emberson, L., Ashmore, M., Mills, G., 2007. Ozone risk assessment for agricultural
600 crops in europe: Further development of stomatal flux and flux–response relationships for european wheat
601 and potato. *Atmospheric Environment* 41 (14), 3022–3040.

602 Pleijel, H., Danielsson, H., Gelang, J., Sild, E., Selldén, G., 1998. Growth stage dependence of the grain yield
603 response to ozone in spring wheat (*triticum aestivum* L.). *Agriculture, ecosystems & environment* 70 (1),
604 61–68.

605 Pleijel, H., Danielsson, H., Ojanperä, K., De Temmerman, L., Högy, P., Badiani, M., Karlsson, P., 2004.
606 Relationships between ozone exposure and yield loss in european wheat and potato—a comparison of
607 concentration-and flux-based exposure indices. *Atmospheric Environment* 38 (15), 2259–2269.

608 Pleijel, H., Danielsson, H., Simpson, D., Mills, G., 2014. Have ozone effects on carbon sequestration been
609 overestimated? a new biomass response function for wheat. *Biogeosciences* 11 (16), 4521–4528.

610 Ratjen, A., Lemaire, G., Kage, H., Plénet, D., Justes, E., 2018. Key variables for simulating leaf area and n
611 status: Biomass based relations versus phenology driven approaches. *European Journal of Agronomy* 100,
612 110–117.

613 Reyenga, P. J., Howden, S. M., Meinke, H., McKeon, G. M., 1999. Modelling global change impacts on wheat
614 cropping in south-east queensland, australia. *Environmental modelling & software* 14 (4), 297–306.

615 Schauburger, B., Rolinski, S., Schaphoff, S., Müller, C., 2019. Global historical soybean and wheat yield loss
616 estimates from ozone pollution considering water and temperature as modifying effects. *Agricultural and*
617 *Forest Meteorology* 265, 1–15.

618 Sharma, A., Ojha, N., Pozzer, A., Beig, G., Gunthe, S. S., 2019. Revisiting the crop yield loss in india at-
619 tributable to ozone. *Atmospheric Environment: X* 1, 100008.

620 Sicard, P., Anav, A., Marco, A. D., Paoletti, E., 2017. Projected global ground-level ozone impacts on vegeta-
621 tion under different emission and climate scenarios. *Atmospheric Chemistry and Physics* 17 (19), 12177–
622 12196.

623 Siddique, K., Kirby, E., Perry, M., 1989. Ear: stem ratio in old and modern wheat varieties; relationship with
624 improvement in number of grains per ear and yield. *Field Crops Research* 21 (1), 59–78.

625 Soja, G., Barnes, J., Posch, M., Vandermeiren, K., Pleijel, H., Mills, G., 2000. Phenological weighting of ozone
626 exposures in the calculation of critical levels for wheat, bean and plantain. *Environmental Pollution* 109 (3),
627 517–524.

628 Tanou, G., Fotopoulos, V., Molassiotis, A., 2012. Priming against environmental challenges and proteomics in
629 plants: update and agricultural perspectives. *Frontiers in Plant Science* 3, 216.

- 630 Temple, P., 1986. Stomatal conductance and transpirational responses of field-grown cotton to ozone. *Plant,*
631 *Cell & Environment* 9 (4), 315–321.
- 632 Tiedemann, A., Pfähler, B., 1994. Growth stage-dependent effects of ozone on the permeability for ions and
633 non-electrolytes of wheat leaves in relation to the susceptibility to septoria nodorum berk. *Physiological and*
634 *molecular plant pathology* 45 (2), 153–167.
- 635 Van Goethem, T., Azevedo, L., Van Zelm, R., Hayes, F., Ashmore, M., Huijbregts, M., 2013. Plant species
636 sensitivity distributions for ozone exposure. *Environmental pollution* 178, 1–6.
- 637 VanLoocke, A., Betzelberger, A. M., Ainsworth, E. A., Bernacchi, C. J., 2012. Rising ozone concentrations
638 decrease soybean evapotranspiration and water use efficiency whilst increasing canopy temperature. *New*
639 *Phytologist* 195 (1), 164–171.
- 640 Wang, W.-N., Cheng, T.-H., Gu, X.-F., Chen, H., Guo, H., Wang, Y., Bao, F.-W., Shi, S.-Y., Xu, B.-R., Zuo, X.,
641 et al., 2017. Assessing spatial and temporal patterns of observed ground-level ozone in china. *Scientific*
642 *reports* 7 (1), 3651.
- 643 Wang, X., Vignjevic, M., Jiang, D., Jacobsen, S., Wollenweber, B., 2014. Improved tolerance to drought
644 stress after anthesis due to priming before anthesis in wheat (*triticum aestivum* L.) var. vinjett. *Journal of*
645 *experimental botany* 65 (22), 6441–6456.
- 646 Wilkinson, S., Mills, G., Illidge, R., Davies, W. J., 2012. How is ozone pollution reducing our food supply?
647 *Journal of Experimental Botany* 63 (2), 527–536.
- 648 Yadav, A., Bhatia, A., Yadav, S., Kumar, V., Singh, B., 2019. The effects of elevated co2 and elevated o3
649 exposure on plant growth, yield and quality of grains of two wheat cultivars grown in north india. *Heliyon*
650 5 (8), e02317.

651 **Appendix A.**

652 *Appendix A.1. SLA function in GLAM-Parti*

653 In Ratjen et al. (2018) the canopy SLA for wheat was expressed as function of LAI (Fig A.10). The
 654 suggested relationship is the following:

$$SLA = \begin{cases} 161.3 + 11.3 \cdot LAI & DVS < 32 \\ 137 + 15.1 \cdot LAI & DVS \geq 32 \end{cases} \quad (A.1)$$

655 For GLAM-Parti, the major limitation of the above piecewise function is the lack of continuity on the first
 656 day when DVS reaches 32 (i.e. Zadoks stage 32). In Fig. A.10, this day is shown in point (LAI1, SLA1). In
 657 order to deal with this discontinuity, the slope and intercept of Eq. A.1 were modified above DVS = 32 as:

$$SLA = \begin{cases} 161.3 + 11.3 \cdot LAI & DVS < 32 \\ z + y \cdot LAI & DVS \geq 32 \end{cases} \quad (A.2)$$

658 where y and z were determined using points (LAI1, SLA1) and (6, 227.6). The second point is the solution of
 659 Eq. A.1 for LAI = 6 and DVS ≥ 32. Based on the two points, y and z were calculated as:

$$y = (SLA1 - 227.6)/(LAI1 - 6) \quad (A.3)$$

660

$$z = 227.6 - 6 \cdot y \quad (A.4)$$

661 Eq. A.2 is used in GLAM-Parti to express SLA as function of LAI and graphical illustration is shown in Fig
 662 A.10 (b).

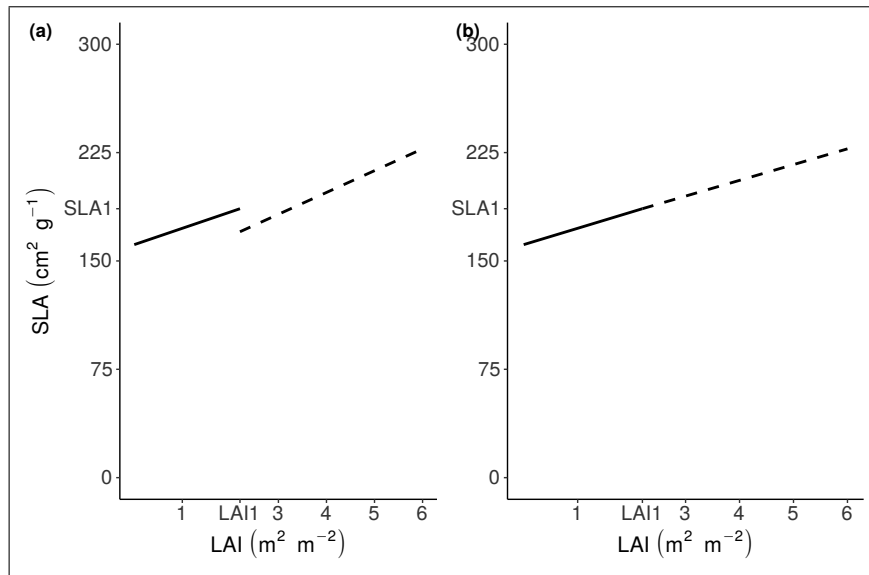


Figure A.10: Canopy SLA as function of LAI (a) in Ratjen et al. (2018) (b) in this study. Point (LAI1, SLA1) is when DVS reaches 32.

663 *Appendix A.2. Expanding GLAM-Parti approach after anthesis*

664 In GLAM-Parti, the above-ground biomass (W_n) consists of leaves, stems, ears and grains as follows:

$$W_n = M_L + M_S + M_E + M_G \quad (\text{A.5})$$

665 where M_L is leaf, M_S is stem, M_E is the non-grain ear mass and M_G is the grain mass. M_E is expressed as
666 function of M_S as follows:

$$M_E = C_E \cdot TT_n / TT_{fl} \cdot M_S \quad (\text{A.6})$$

667 where TT_n is the thermal time elapsed from terminal spikelet initiation (TS) until day n after TS and TT_{fl} is the
668 thermal time requirement from TS to anthesis. C_E expresses the ratio of ear: stem mass at anthesis, which
669 is set to 0.5 for modern wheat varieties (Siddique et al., 1989). M_G is expressed as function of W_n using the
670 harvest index (HI) as follows:

$$M_G = HI \cdot W_n \quad (\text{A.7})$$

671 Eq. A.6, A.7 can be combined to describe Eq. A.5 as:

$$W_n = (1/(1 - HI)) \cdot (M_L + (1 + C_E \cdot TT_n / TT_{fl}) \cdot M_S) \quad (\text{A.8})$$

672 Eq. A.8 can be further manipulated to express W_n as function of leaf area change. M_L is expressed as:

$$M_L = LAI / SLA + M_{YL} \quad (\text{A.9})$$

673 where LAI is green leaf area index, SLA is canopy specific leaf area and M_{YL} is the mass of yellow leaves.
674 LAI can be expanded as:

$$LAI_n = LAI_{n-1} + dL \quad (\text{A.10})$$

675 where LAI_n is the value of LAI at any given n day, LAI_{n-1} is LAI of the previous day and dL is the leaf area
676 change between the two consecutive days. The mass of stems (M_S) is expressed with an allometric relation-
677 ship according to M_L as:

$$M_S = h \cdot M_L^g \quad (\text{A.11})$$

678 where g , h are allometric coefficients. Eq. A.2, A.9, A.10, A.11 can be combined to express Eq. A.8 as:

$$W_n = (1/(1 - HI)) \cdot \left(\frac{LAI_{n-1} + dL}{z + y \cdot (LAI_{n-1} + dL)} + M_{YL} + (1 + C_E \cdot TT_n / TT_{fl}) \cdot h \left(\frac{LAI_{n-1} + dL}{z + y \cdot (LAI_{n-1} + dL)} + M_{YL} \right)^g \right) \quad (\text{A.12})$$

679 where the slope and intercept of Eq. A.2 (y , z) vary before and after DVS = 32. Eq. A.12 expresses W_n as
680 function of the leaf area change (dL) and is used for the implementation of the SEMAC methodology during

681 the full crop cycle in GLAM-Parti. A detailed description of the SEMAC approach is given in Droutsas et al.
 682 (2019).

683 Moreover, the model was parameterized to account for the canopy leaf mass loss which mainly occurs
 684 during the period of rapid leaf senescence after anthesis. Whenever a negative value of leaf area change (dL)
 685 was estimated, the mass of yellow leaves (M_{YL}) was updated as:

$$M_{YL(n)} = M_{YL(n-1)} + C_{yl} * (|dL|/SLA) \quad (\text{A.13})$$

686 where $M_{YL(n)}$ is the mass of yellow leaves on the n day of the crop cycle, $M_{YL(n-1)}$ is the mass of yellow leaves
 687 on the previous day (n-1), SLA is the canopy specific leaf area and C_{yl} is the ratio of yellow:green leaf mass
 688 which was set to 0.68 to account for the leaf mass loss due to the remobilization of dry mass (Borrell et al.,
 689 1989).

690 Appendix A.3. O_3 effect on evapotranspiration in GLAM-ROC

691 The statistical formula for the expression of percentage change in cumulative evapotranspiration (pCET)
 692 according to effective AOT40 (efAOT40) is given below (Fig. 3 (b)):

$$pCET = -0.021 + 0.018 \cdot efAOT40 - 0.000356 \cdot efAOT40^2 \quad (\text{A.14})$$

693 where at any given n day of the growing season, pCET between the Control and O_3 treatments is defined as:

$$pCET_n = \frac{CET_{AA_n} - CET_{oz_n}}{CET_{oz_n}} = \frac{CET_{AA_n}}{CET_{oz_n}} - 1 = \frac{CET_{AA_n}}{CET_{oz_{n-1}} + ET_{oz_n}} - 1 \quad (\text{A.15})$$

694 where CET_{AA} and CET_{oz} are the cumulative evapotranspiration of the control and O_3 treatment respectively.

695 The substitution of Eq. A.15 into A.14 and solving for ET_{oz} gives:

$$ET_{oz_n} = \frac{CET_{AA_n}}{0.979 + 0.018 \cdot efAOT40 - 0.000365 \cdot efAOT40^2} - CET_{oz_{n-1}} \quad (\text{A.16})$$

696 Eq. A.16 is used in GLAM-ROC to calculate the O_3 -induced decrease in ET (ET_{oz}) in comparison with the
 697 same plant growing under optimal conditions.

TE reduction factor		dHI/dt reduction factor	
c1	d1	c2	d2
-0.0029	1.029	-0.00145	1.0145

Table A.5: Slope and intercept of O₃-induced reduction in transpiration efficiency (TE) and rate of change in harvest index (dHI/dt) relative to Control.

Parameter	Unit	Range	GLAM-ROC value	Source
E _{TN,max}	g kg ⁻¹	[3 - 10.6] ^a	9.3	Christy et al. (2018)
dHI/dt	day ⁻¹	[0.0064 - 0.0137]	0.0135	Moot et al. (1996)

Table A.6: Values and units of GLAM-ROC calibrated parameters.

^a Average CO₂ concentration in the chambers was around 530 ppm (see Table 1). The upper boundary of E_{TN,max} was multiplied by 1.18 to account for the CO₂ fertilization effect on TE (Reyenga et al., 1999).