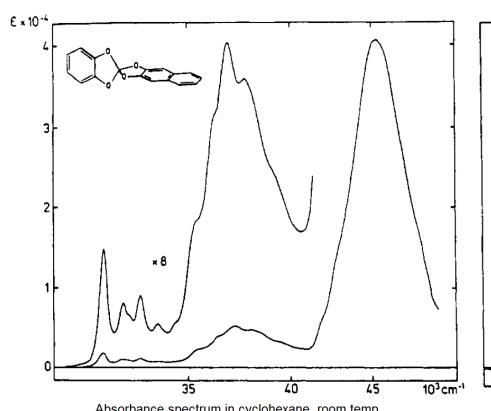


Structural Applications of Polarization Spectroscopy on Aligned Molecular Assemblies

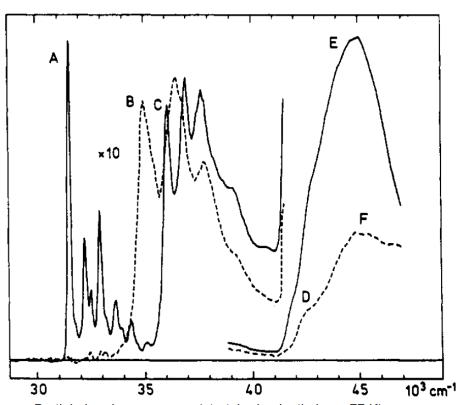
Methods in Soft Materials Science RUC, January 29, 2015

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Optical absorbance spectroscopy:



Absorbance spectrum in cyclohexane, room temp.



Partial absorbance curves (stretched polyethylene, 77 K): Full and dashed curves indicate long- and short-axis polarized absorbance

Linear Dichroism (LD)

Programme:

1) LD spectroscopy: Working equations for uniaxial samples (quinizarin)

Molecular structural applications:

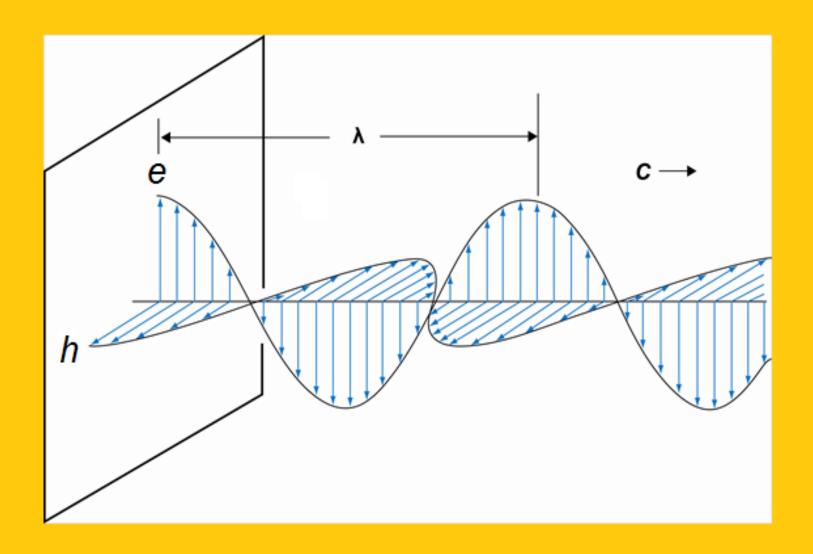
- 2) 1,6,6aλ⁴-Trithiapentalene
- 3) Diphenyl-diacetylene
- 4) Ellagic Acid + DNA (Flow LD)

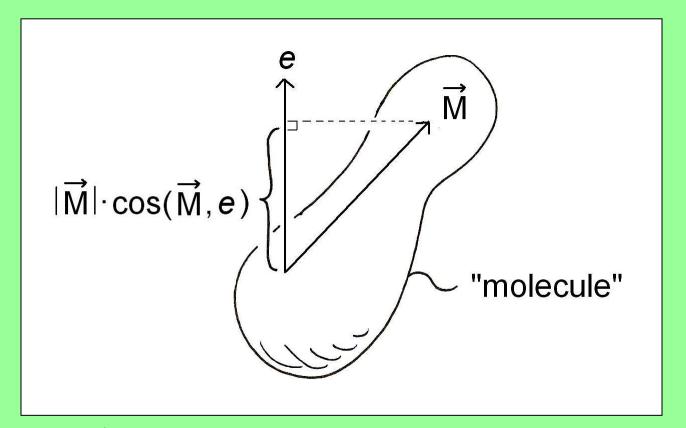
Main characteristica of optical absorption bands:

- 1) Transition energy (frequency, wavenumber, wavelength)
- 2) Intensity (absorbance, oscillator strength)

3) Polarization, i.e., transition moment direction

Plane Polarized Radiation

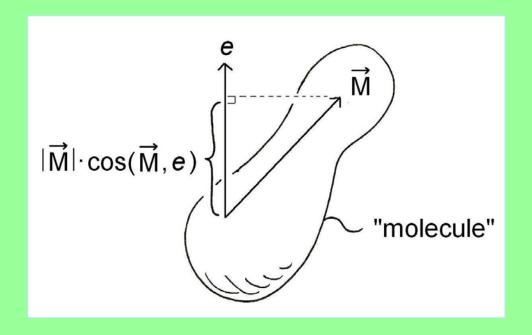




 \vec{M} = transition moment vector e = electric field vector (\vec{M}, e) = angle between \vec{M} and e

Transition Probability $\propto |\vec{M}|^2 \cos^2(\vec{M}, e)$

Absorbance Measurement



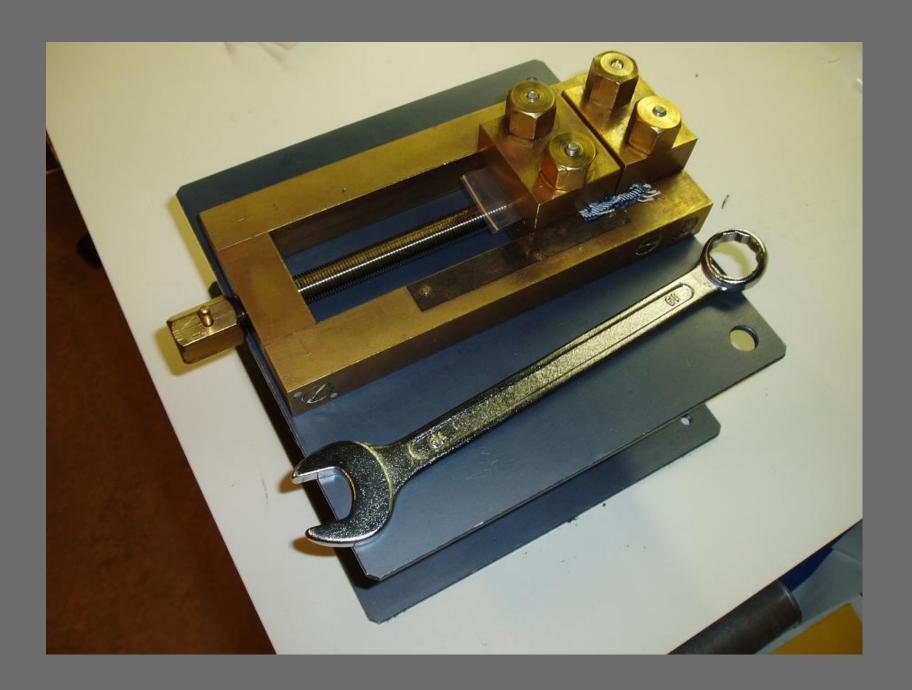
Absorbance =
$$A(\widetilde{\nu}) \cdot \langle \cos^2(\vec{M}, e) \rangle$$

where $A(\tilde{v})$ is the absorbance measured for $(\vec{M}, e) = 0$

Stretched Polyethylene as an Anisotropic Solvent

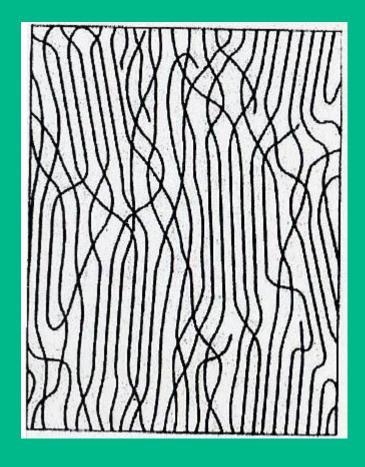


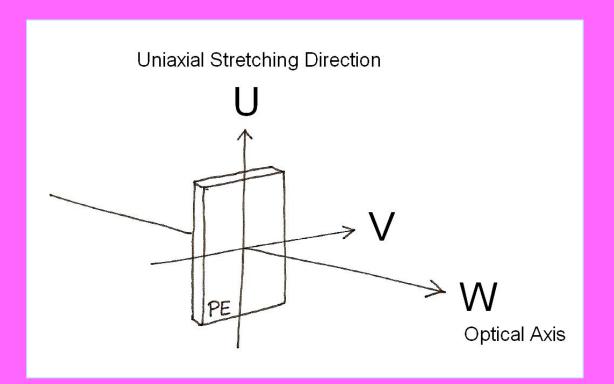




Simplified model of polyethylene (PE)

Corresponding model of stretched polyethylene (PE)





$$\mathsf{E}_\mathsf{U}(\widetilde{\nu}) = \mathsf{A}(\widetilde{\nu}) \cdot \left\langle \mathsf{cos}^2(\vec{\mathsf{M}},\mathsf{U}) \right\rangle$$

$$\mathsf{E}_{\mathsf{V}}(\widetilde{\nu}) = \mathsf{A}(\widetilde{\nu}) \cdot \left\langle \mathsf{cos}^{2}(\vec{\mathsf{M}}, \mathsf{V}) \right\rangle$$

$$\frac{\mathsf{E}_{\mathsf{W}}(\widetilde{\nu}) = \mathsf{A}(\widetilde{\nu}) \cdot \left\langle \mathsf{cos}^{2}(\vec{\mathsf{M}}, \mathsf{W}) \right\rangle}{\mathsf{Cos}^{2}(\vec{\mathsf{M}}, \mathsf{W})}$$

Uniaxial Sample

$$\begin{split} &\left\langle cos^{2}(\vec{M},V)\right\rangle =\left\langle cos^{2}(\vec{M},W)\right\rangle \\ &\left\langle cos^{2}(\vec{M},U)\right\rangle +\left\langle cos^{2}(\vec{M},V)\right\rangle +\left\langle cos^{2}(\vec{M},W)\right\rangle =1 \end{split}$$

$$\Rightarrow \left\langle \cos^2(\vec{M}, V) \right\rangle = \frac{1}{2} \left(1 - \left\langle \cos^2(\vec{M}, U) \right\rangle \right)$$

We define the Orientation Factor *K*:

$$K \equiv \left\langle \cos^2(\vec{M}, U) \right\rangle$$

Thulstrup, Eggers, Michl 1970

Uniaxial Sample

$$K \equiv \left\langle \cos^2(\vec{M}, U) \right\rangle$$

$$\mathsf{E}_\mathsf{U}(\widetilde{\nu}) = \mathsf{A}(\widetilde{\nu}) \cdot \left\langle \mathsf{cos}^2(\vec{\mathsf{M}},\mathsf{U}) \right\rangle = \mathsf{A}(\widetilde{\nu}) \cdot \mathsf{K}$$

$$\mathsf{E}_{\mathsf{V}}(\widetilde{\mathsf{v}}) = \mathsf{A}(\widetilde{\mathsf{v}}) \cdot \left\langle \mathsf{cos}^{2}(\vec{\mathsf{M}}, \mathsf{V}) \right\rangle = \mathsf{A}(\widetilde{\mathsf{v}}) \cdot \frac{1}{2} (1 - K)$$

Dichroic Ratio, d:

$$d = \frac{\mathsf{E}_{\mathsf{U}}(\widetilde{v})}{\mathsf{E}_{\mathsf{V}}(\widetilde{v})} = \frac{2K}{1-K} \implies K = \frac{d}{2+d}$$

In general, several transitions "i" contribute to the observed dichroic absorption curves:

$$E_{U}(\widetilde{v}) = \sum_{i} A_{i}(\widetilde{v}) \cdot K_{i}$$

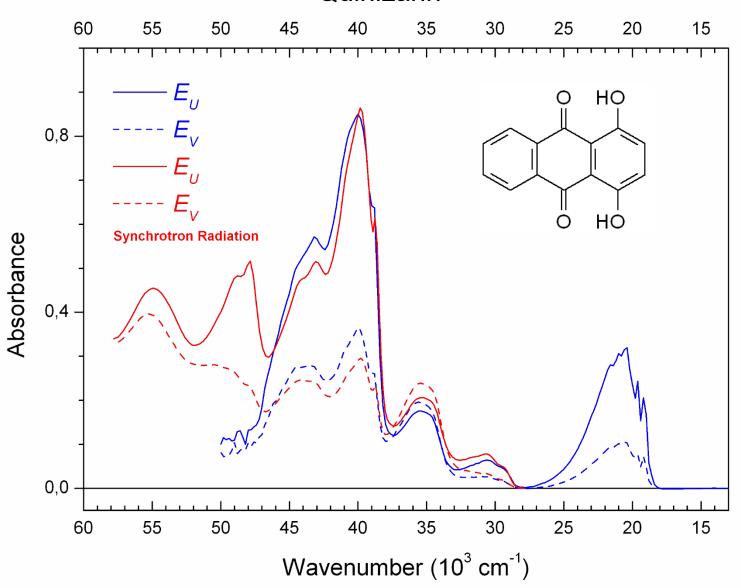
$$E_{V}(\widetilde{v}) = \sum_{i} A_{i}(\widetilde{v}) \cdot \frac{1}{2} (1 - K_{i})$$

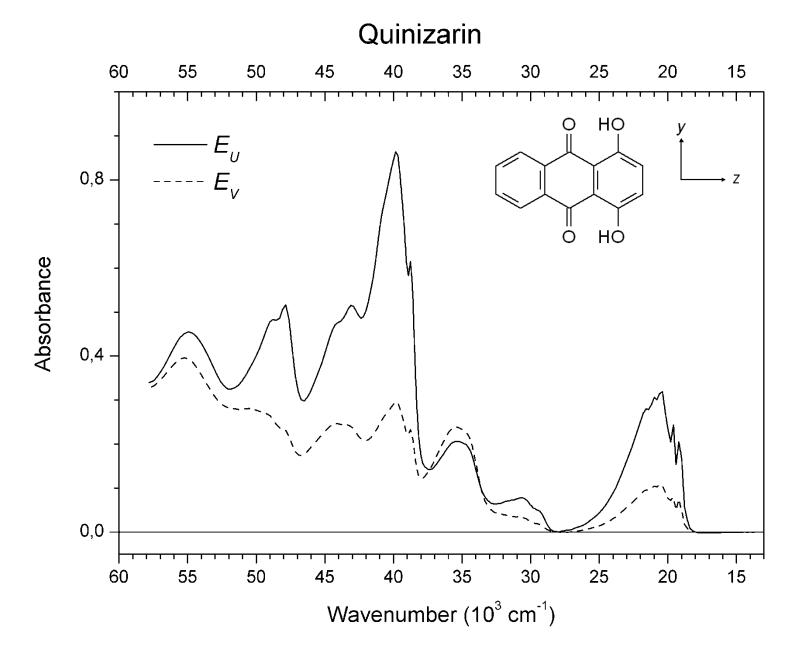
TEM procedure for overlapping transitions:

$$r_{\kappa} = (1 - K) \cdot E_{\cup}(\widetilde{\nu}) - 2K \cdot E_{\vee}(\widetilde{\nu})$$

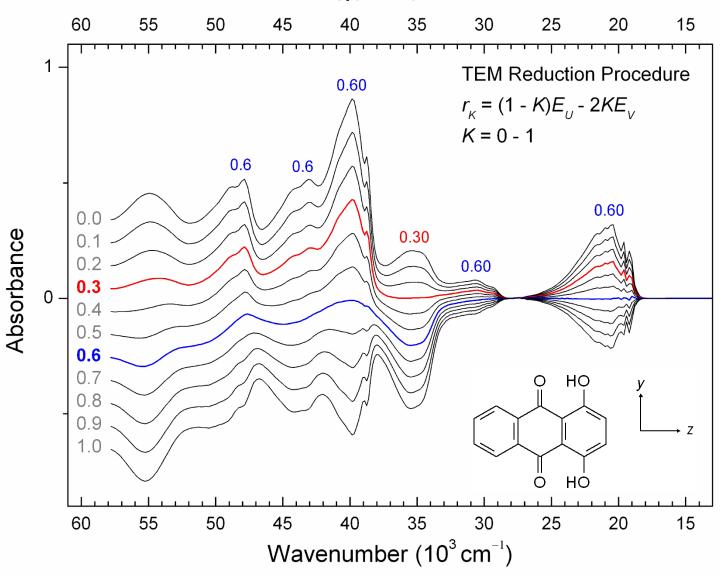
A spectral feature (peak or shoulder) due to transition i vanishes from r_K when $K = K_i$. The K_i value can thus be determined by visual inspection.

Quinizarin

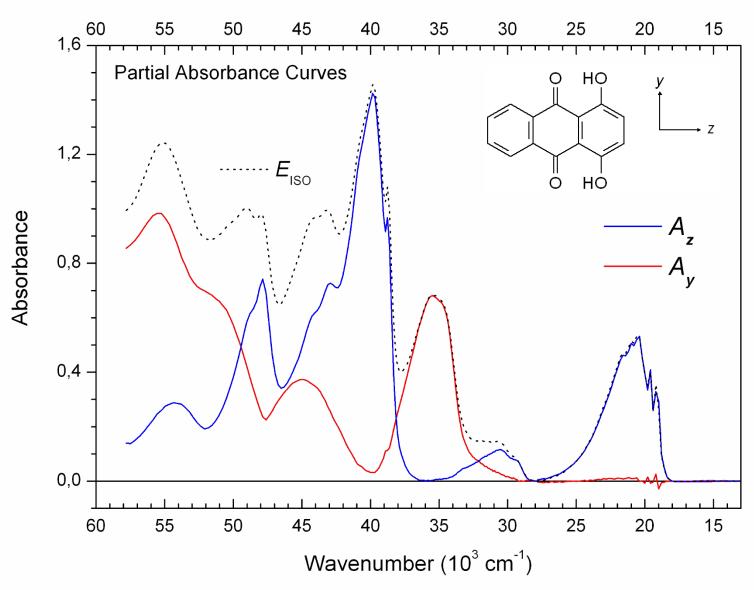


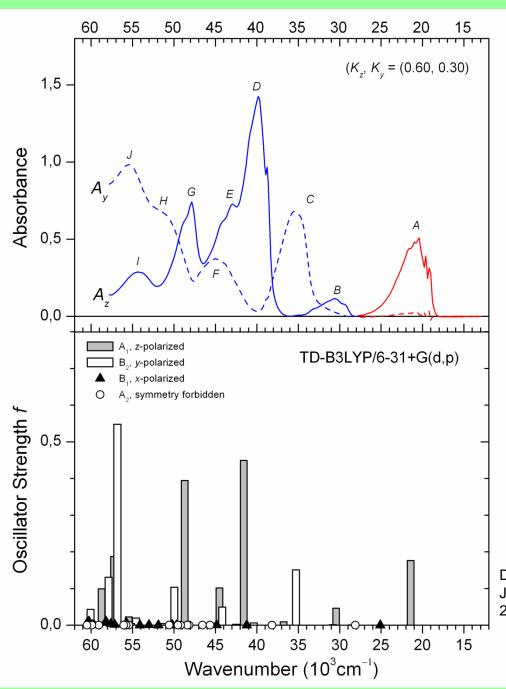






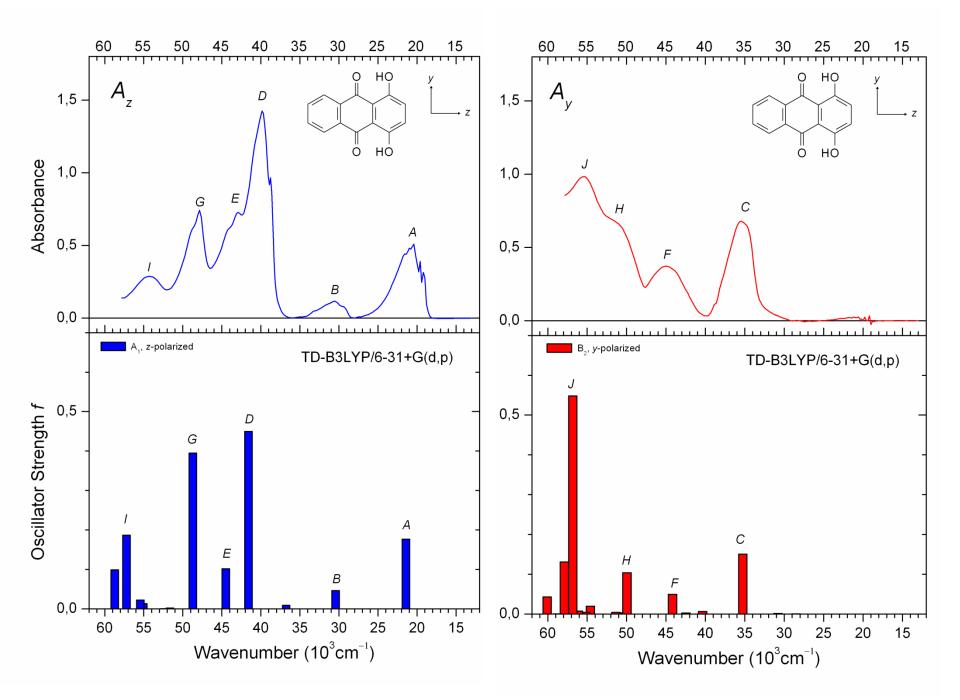
Quinizarin



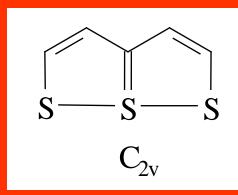


Quinizarin

D.D. Nguyen, N.C. Jones, S.V. Hoffmann, J. Spanget-Larsen, *Spectrochim. Acta A* **77**, 279-286 (2010).



1,6,6aλ⁴-Trithiapentalene



If a molecule has D_{2h} , D_2 , or C_{2v} point group symmetry, optically allowed transitions must be polarized along one of the three mutually perpendicular molecular symmetry axes x, y, and z.

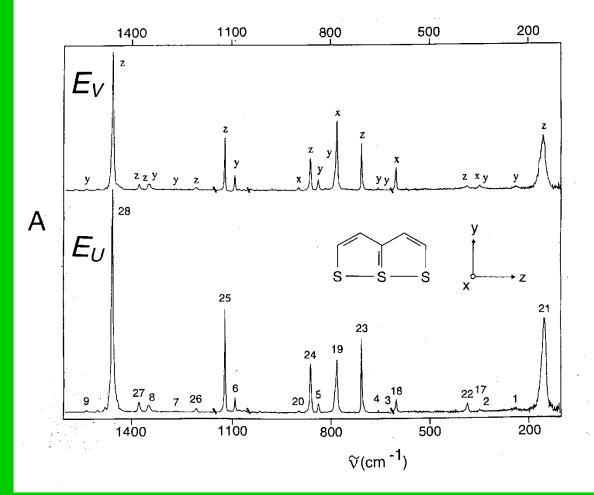
The observed K values should thus adopt only three distinct values, corresponding to the orientation factors K_x , K_y , and K_z for the three symmetry axes, and the three values should add up to unity:

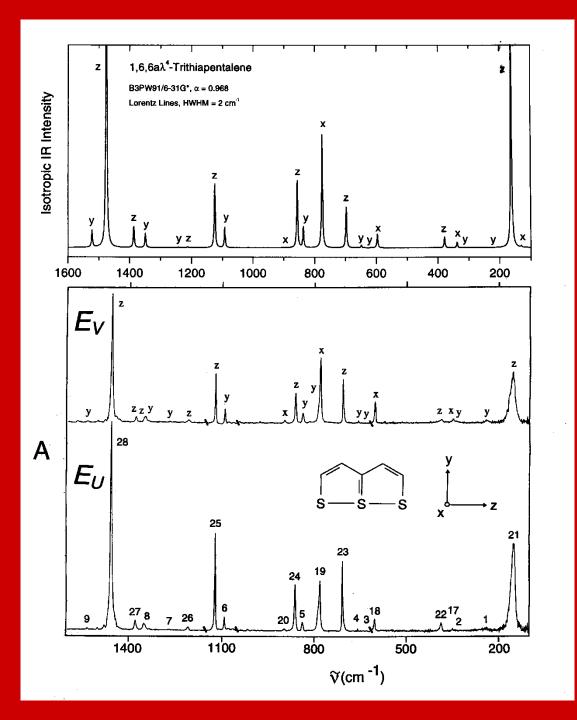
$$K_x + K_y + K_z = 1$$

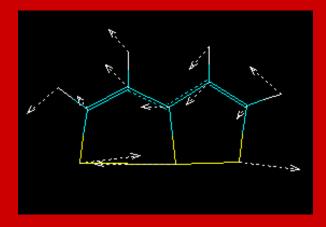
- (1) If more than three significantly different K values are observed, the molecular symmetry must be different from D_{2h} , D_2 , and C_{2v} .
- (2) If the observed K values for a large number of transitions fall in three distinct groups, with the three characteristic values adding up to unity, this is strong evidence for the presence of molecular symmetry elements, consistent with D_{2h} , D_2 , or C_{2v} .

$$K_i \equiv \left\langle \cos^2(\vec{M}_i, U) \right\rangle$$

$$(K_x, K_y, K_z) = (0.240 \pm 0.01, 0.306 \pm 0.01, 0.462 \pm 0.02)$$







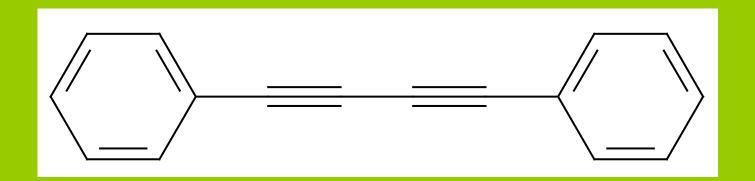
"Bell-Clapper" mode (153 cm⁻¹)

The observed orientation factors Indicate that the molecule has a rigid, symmetrical structure (C_{2v})

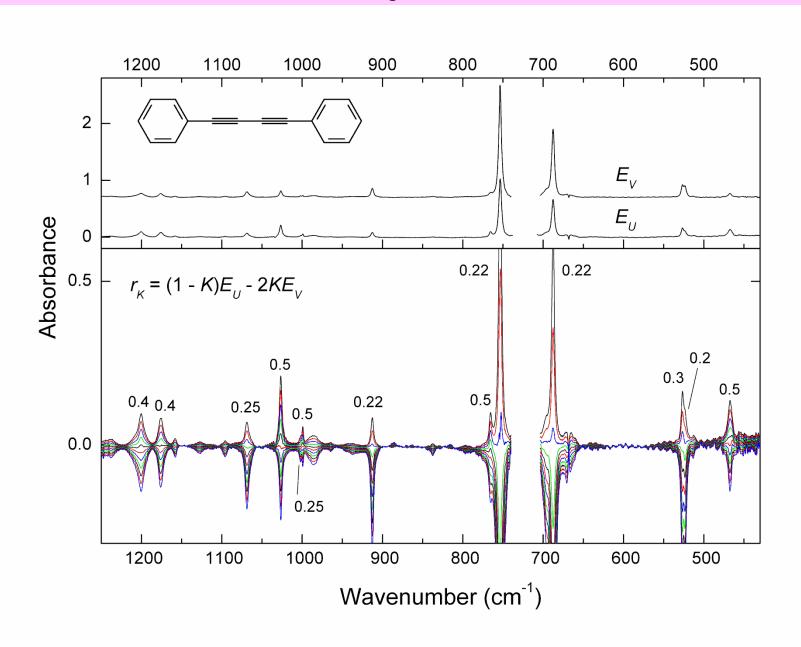
Experimental observation of the "Bell-Clapper" transition confirms it

K.B. Andersen, J. Abildgaard, J.G. Radziszewski, J. Spanget-Larsen: *J. Phys. Chem. A* **101**, 4475 (1997)

Diphenyldiacetylene (DPDA)



FTIR-LD of DPDA aligned in stretched LDPE

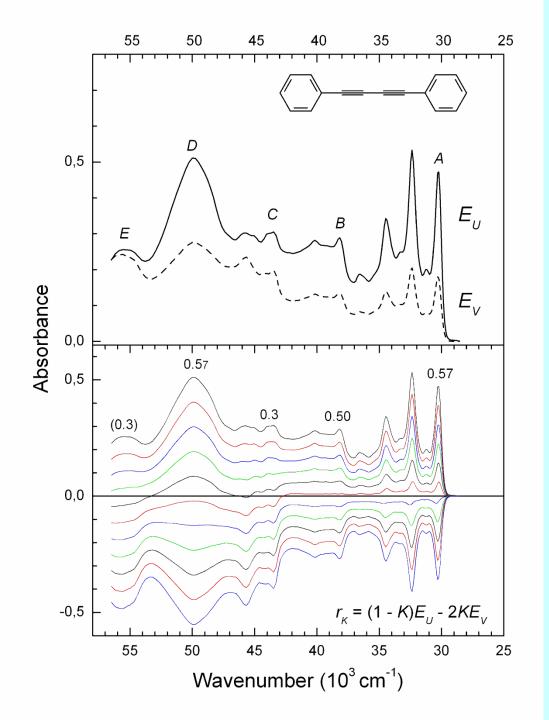


UV-SRLD of DPDA aligned in LDPE

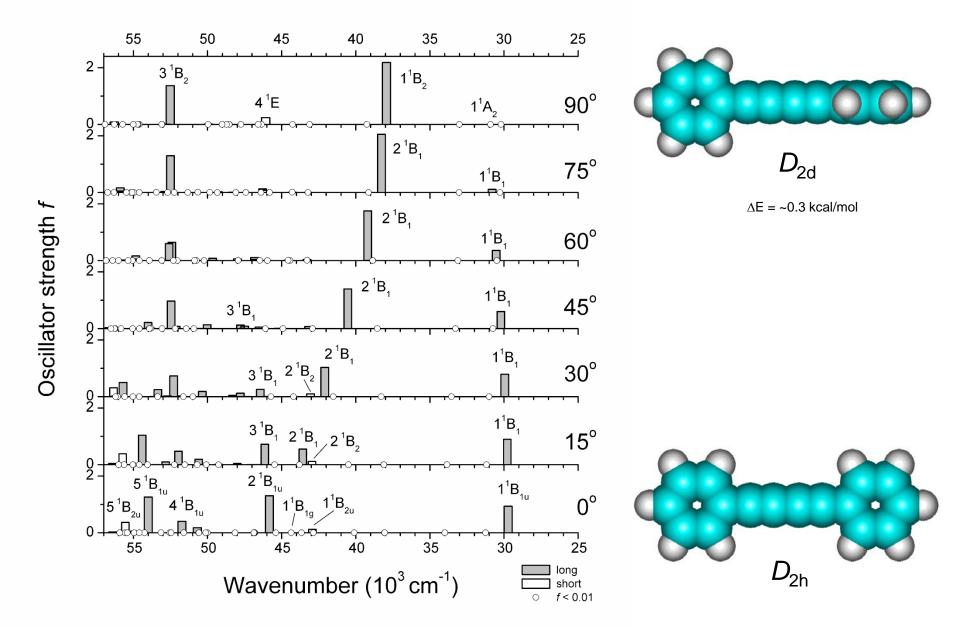
The observed orientation factors are inconsistent with the presence of a single, symmetric species.

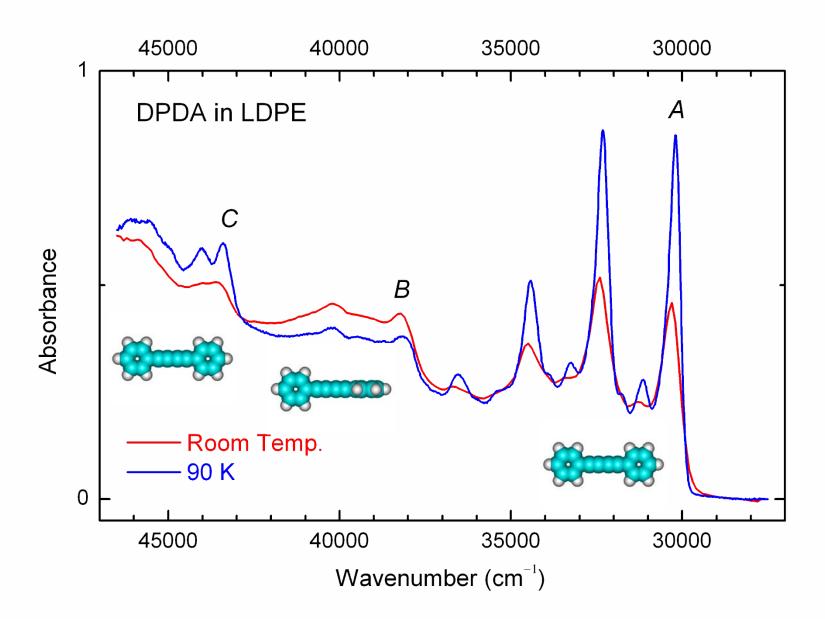
Do we observe overlapping contributions from different rotamers?

P.W. Thulstrup, S.V. Hoffmann, B.K.V. Hansen, J. Spanget-Larsen, *PCCP* **13**, 16168-16174 (2011)



TD-PBE1PBE/6-31+G*//6-31G*





BPEA

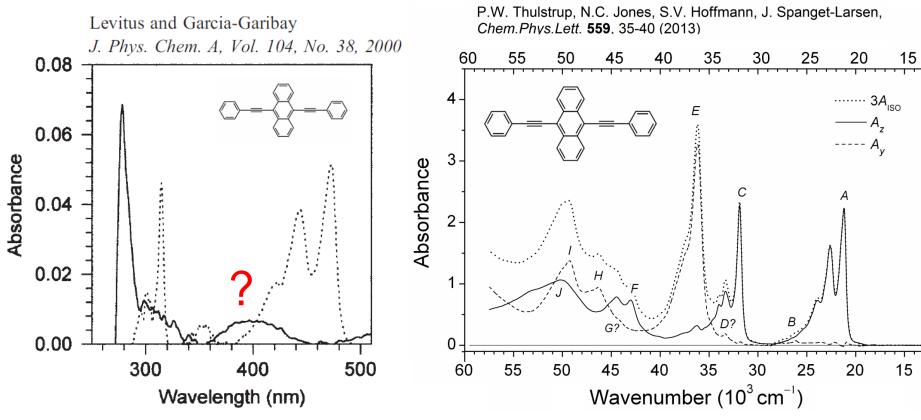
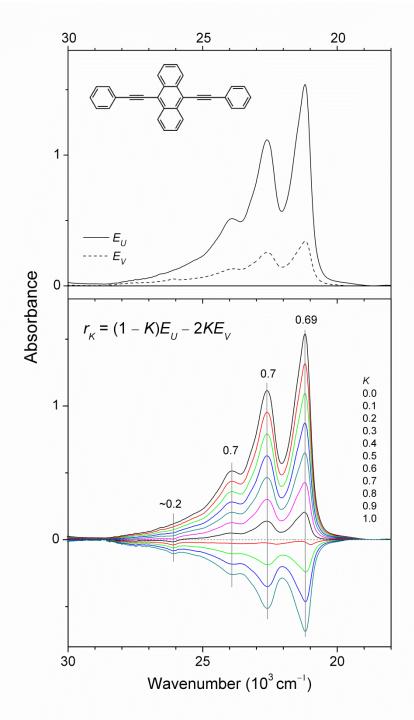


Figure 4. Purely polarized spectra obtained from the stepwise reduction procedure discussed in the text. Solid line: A_y , spectrum polarized along the *y*-axis. Dotted line: A_z , spectrum polarized along the *z*-axis.

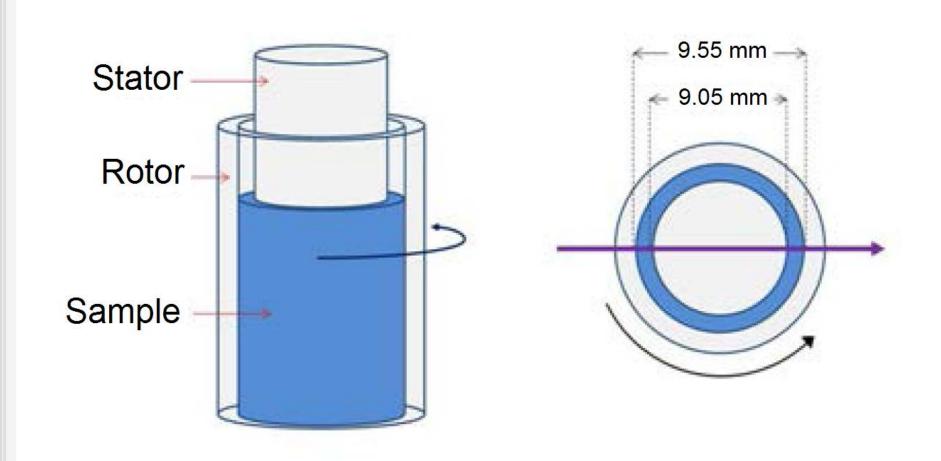


Flow LD Experiments

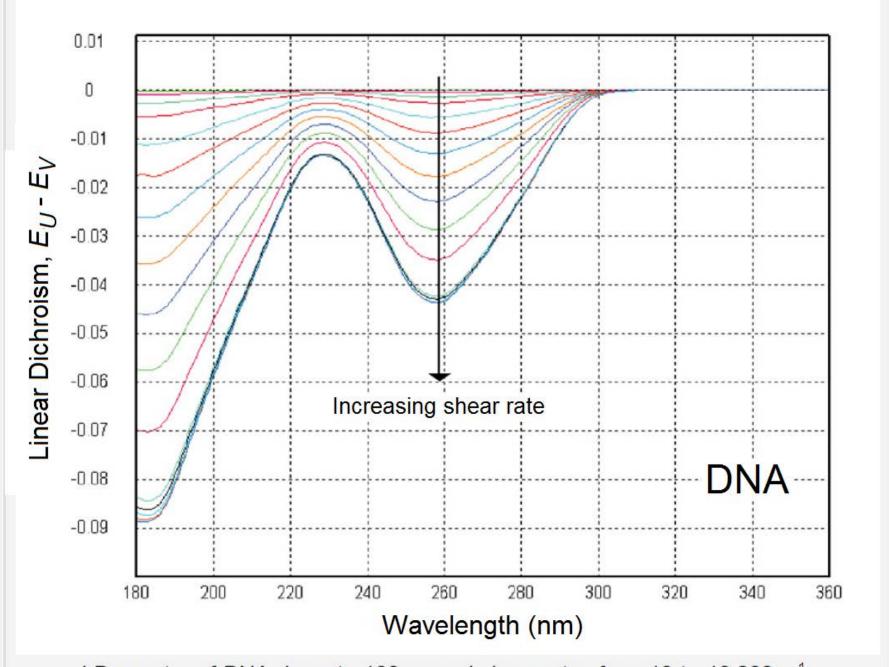
Biological Polymers

One way to obtain an aligned sample of a biological polymer is by application of a flow gradient. For example, macromolecular DNA is oriented in a flow gradient, whereas small molecules are not.

Hence, Flow LD experiments can be applied in investigations of the interaction between small molecules and DNA.



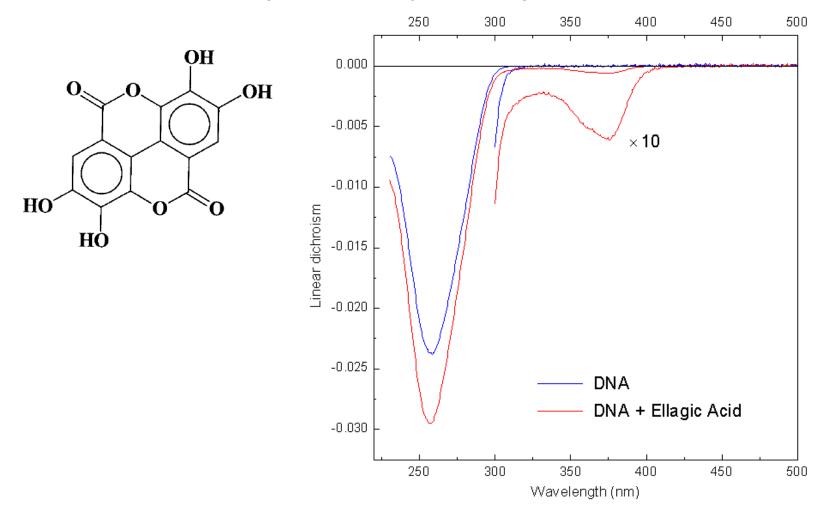
Schematics of the Couette Cell



LD spectra of DNA down to 180nm and shear rates from 10 to 10,200 s⁻¹

Interaction between Ellagic Acid and Calf Thymus DNA Studied with Flow Linear Dichroism UV-VIS Spectroscopy¹

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Acknowledgements

RUC:

Nguyen Duc Duy Bjarke K. V. Hansen Eva M. Karlsen



ISA:

Søren Vrønning Hoffmann Nykola C. Jones



KU Life:

Peter Waaben Thulstrup



