



# Gasification biochar as a valuable by-product for carbon sequestration and soil amendment

Hansen, Veronika; Müller-Stöver, Dorette; Ahrenfeldt, Jesper; Holm, Jens Kai; Henriksen, Ulrik Birk; Hauggaard-Nielsen, Henrik

Published in: **Biomass & Bioenergy** 

DOI: 10.1016/j.biombioe.2014.10.013

Publication date: 2015

Document Version Peer reviewed version

Citation for published version (APA):

Hansen, V., Müller-Stöver, D., Ahrenfeldt, J., Holm, J. K., Henriksen, U. B., & Hauggaard-Nielsen, H. (2015). Gasification biochar as a valuable by-product for carbon sequestration and soil amendment. *Biomass & Bioenergy*, 72, 300-308. https://doi.org/10.1016/j.biombioe.2014.10.013

**General rights** 

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
  You may not further distribute the material or use it for any profit-making activity or commercial gain.
  You may freely distribute the URL identifying the publication in the public portal.

#### Take down policy

If you believe that this document breaches copyright please contact rucforsk@kb.dk providing details, and we will remove access to the work immediately and investigate your claim.

Elsevier Editorial System(tm) for Biomass and Bioenergy Manuscript Draft

#### Manuscript Number: JBB-D-14-00319R3

Title: Gasification biochar as a valuable by-product for carbon sequestration and soil amendment

Article Type: Research Paper

Keywords: Gasification, Bioenergy efficiency, Biochar soil amendment, Carbon sequestration, Soil quality improvement

Corresponding Author: Mrs. Veronika Hansen,

Corresponding Author's Institution: Technical University of Denmark

First Author: Veronika Hansen

Order of Authors: Veronika Hansen; Dorette Müller-Stöver; Jesper Ahrenfeldt; Jens Kai Holm; Ulrik B Henriksen; Henrik Hauggaard-Nielsen

Abstract: Thermal gasification of various biomass residues is a promising technology for combining bioenergy production with soil fertility management through the application of the resulting biochar as soil amendment. In this study, we investigated gasification biochar (GB) materials originating from two major global biomass fuels: straw gasification biochar (SGB) and wood gasification biochar (WGB), produced by a Low Temperature Circulating Fluidized Bed gasifier (LT-CFB) and a TwoStage gasifier, respectively, optimized to energy conversion. Stability of carbon in GB against microbial degradation was assessed in a short-term soil incubation study and compared to the traditional practice of direct incorporation of cereal straw. The GBs were chemically and physically characterized to evaluate their potential to improve soil quality parameters. After 110 days of incubation, about 3 % of the added GB carbon was respired as CO2, compared to 80 % of the straw carbon added. The stability of GB was also confirmed by low H/C and O/C ratios with lowest values for WGB (H/C 0.01 and O/C 0.14). The soil application of GBs exhibited a liming effect increasing the soil pH from ca 8 to 9. Results from scanning electron microscopy and BET analyses showed high porosity and specific surface area of both GBs, indicating a high potential to increase important soil quality parameters such as soil structure, nutrient and water retention especially for WGB. These results seem promising regarding the possibility to combine an efficient bioenergy production with various soil aspects such as carbon sequestration and soil quality improvements.

Response to Reviewers: Response to Reviewers' comments: The units and ratios were corrected (L 143, L184). Treatments description were also improved (L159-64) The references have been corrected.

Thank you for your comments. Best regards, Veronika Hansen

20.03.2014

Submission of manuscript titled:

### Gasification biochar as valuable by-product for carbon sequestration and soil amendment

This manuscript demonstrates the potential for combining of bioenergy production and residual biochar application as soil improving and carbon sequestration agent. This study shows that carbon in gasification biochar is stable against microbial degradation. Furthermore, the liming effect, high porosity and specific surface area of the gasification biochar indicate the ability of the biochar to improve important soil quality parameters such as structure, water and nutrient retention.

Therefore, gasification of crop residues and wood waste is a promising way of producing sustainable bioenergy and reaching the political goals of fossil fuel free society, and at the same time sustaining or even improving of the soil quality, which is crucial for meeting the increasing demand for producing food and energy crops.

We hope that our experimental study might be interesting and relevant for publication in Biomass and Bioenergy. We are looking forward to receive the reviewer's comments and evaluation.

Best regards,

Veronika Hansen PhD student Technical university of Denmark Department of Chemical and Biochemical Engineering +45 20 62 05 22 veha@kt.dtu.dk Respons to Reviewers' comments: The units and ratios were corrected (L 143, L184). Treatments description were also improved (L159-64)

The references have been corrected.

Thank you for your comments.

Best regards,

Veronika Hansen

# Highlights

- Biomass gasification has a potential to combine the efficient production of bioenergy with valuable biochar residuals for soil quality improvements.
- The two investigated gasification biochars are recalcitrant indicating soil carbon sequestration potential.
- Gasification biochars have a potential as soil improvers due to high specific surface area, porosity, liming effect and low PAH content.

# 1 Gasification biochar as a valuable by-product for carbon sequestration and soil amendment

- 2 Veronika Hansen<sup>a,\*</sup>, Dorette Müller-Stöver<sup>a</sup>, Jesper Ahrenfeldt<sup>a</sup>, Jens Kai Holm<sup>b</sup>, Ulrik Birk
- 3 Henriksen<sup>a</sup>, Henrik Hauggaard-Nielsen<sup>c</sup>
- <sup>4</sup> <sup>a</sup> Department of Chemical and Biochemical Engineering, Technical University of Denmark,
- 5 Frederiksborgvej 399, 4000 Roskilde, Denmark.
- <sup>6</sup> <sup>b</sup> DONG Energy Thermal Power A/S, Nesa Allé 1, 2820 Gentofte, Denmark.
- <sup>c</sup> Department of Environmental, Social and Spatial Change, Roskilde University, 4000 Roskilde,
- 8 Denmark.
- 9 \*Corresponding author: Tel.: +45 20 62 05 22. E-mail address: <u>veha@kt.dtu.dk</u>(V. Hansen)

#### 11 Abstract

12 Thermal gasification of various biomass residues is a promising technology for combining 13 bioenergy production with soil fertility management through the application of the resulting biochar 14 as soil amendment. In this study, we investigated gasification biochar (GB) materials originating 15 from two major global biomass fuels: straw gasification biochar (SGB) and wood gasification 16 biochar (WGB), produced by a Low Temperature Circulating Fluidized Bed gasifier (LT-CFB) and 17 a TwoStage gasifier, respectively, optimized to energy conversion. Stability of carbon in GB against 18 microbial degradation was assessed in a short-term soil incubation study and compared to the 19 traditional practice of direct incorporation of cereal straw. The GBs were chemically and physically 20 characterized to evaluate their potential to improve soil quality parameters. After 110 days of 21 incubation, about 3 % of the added GB carbon was respired as CO<sub>2</sub>, compared to 80 % of the straw 22 carbon added. The stability of GB was also confirmed by low H/C and O/C atomic ratios with 23 lowest values for WGB (H/C 0.01 and O/C 0.14). The soil application of GBs exhibited a liming 24 effect increasing the soil pH from ca 8 to 9. Results from scanning electron microscopy and BET 25 analyses showed high porosity and specific surface area of both GBs, indicating a high potential to increase important soil quality parameters such as soil structure, nutrient and water retention 26 27 especially for WGB. These results seem promising regarding the possibility to combine an efficient 28 bioenergy production with various soil aspects such as carbon sequestration and soil quality 29 improvements.

30

# 31 Keywords

Gasification, Bioenergy efficiency, Biochar soil amendment, Carbon sequestration, Soil quality
 improvement

#### 34 **1. Introduction**

35 Biomass gasification for combined heat and power (CHP) production has the potential to become an 36 efficient and flexible way to generate bioenergy, as a broad variety of biomass residues and other 37 organic resources can be utilized [1, 2]. In Denmark effective gasification platforms for the two major global biomass fuels, wood chips and cereal straw, are currently scaled up and close to 38 39 commercial application: (1) Low Temperature Circulating Fluidized Bed gasifier (LT-CFB), 40 specifically designed to produce energy from biomasses with high ash contents (such as straw) and 41 (2) TwoStage gasifier, designed for converting woody biomass. The LT-CFB technology has been 42 demonstrated in continuous operation, as a 6 MW demonstration plant, and the first 2 MW 43 commercial plant for continues CHP production with the TwoStage process is about to produce 44 power and district heating for a local community, Hilleroed Municipality, Denmark. This plant will 45 produce approximately 64 tons of biochar residues annually, while the planned 60 MW full scale 46 commercial LT-CFB plant is going to generate approximately 10 000 tons of carbon-rich residues 47 per year. The potential further upscaling and expanding of those processes requires a strategy for 48 the sustainable utilization of a growing amount of biochar residues produced. Recirculation and 49 utilization of those residues to agricultural land, instead of costly disposing as a waste, would 50 improve the sustainability and economy of the bioenergy production. Gasification biochar generally 51 contains a considerable amount of minerals and recalcitrant carbon and is considered an attractive 52 product for soil amendment due to its fertilizer and carbon sequestration potential [3, 4].

53 Carbon sequestration in soil mitigates the effect of climate change [5], and may furthermore help to 54 maintain or even improve the soil fertility. This is of key importance to be able to fulfil the 55 increasing global demand for producing crops for both food and energy [6]. Soil organic carbon 56 (SOC) influences the physical, chemical and biological properties of the soil, and is essential for 57 good soil quality [7]. Increasing SOC has been shown to improve soil aggregation, water 58 infiltration, and water and nutrient retention [8, 9]. Traditional annual incorporation of crop residues 59 such as cereal straw can increase soil organic matter content [10], therefore there is a concern that 60 the removal of residues from the field for energy production may lead to soil degradation [11]. 61 Gasification of biomass and returning the residual biochar-carbon to the field is regarded as a promising strategy combining effective bioenergy generation with the maintenance of soil carbon 62 63 stocks [2]. Utilizing low quality wood and residues from timber harvesting for bioenergy production 64 and subsequent addition of wood biochar to agricultural soils may be another strategy to increase 65 SOC and improve arable soils' productivity, creating novel synergies between the agricultural and 66 forestry sectors. Nevertheless, since there are qualitative differences in the molecular structure of 67 pyrogenic carbon compared to the stable carbon derived from microbial/enzymatic soil processes 68 [12], the impacts of substituting crop residue incorporation with the addition of gasification biochar 69 (GB) on soil services are largely unknown and should be thoroughly investigated before 70 implementing this into practice [8].

71 Several studies have shown positive impacts of pyrolysis biochar, produced at relatively low 72 temperatures  $(400 - 600^{\circ}C)$ , on soil properties [13, 14], which are, however, highly dependent on 73 biochar feedstock and thermal processing conditions [15]. The physical properties of biochars, such 74 as high porosity and specific surface area (BET), may result in an increase of not only soil water 75 retention [16], water infiltration, and cation exchange capacity [5, 13], but also soil microbial 76 activity [14]. Chemical properties, such as low hydrogen-to-carbon (H/C) and oxygen-to-carbon 77 (O/C) ratios, result in high stability of biochar against microbial degradation in soil [17]. Compared 78 to pyrolysis biochar, GB is produced at higher temperatures (around 700 - 1100°C), using low 79 amounts of oxygen. Gasification results in higher energy yields compared to pyrolysis and leaves 80 biochar with less, but more stable carbon, compared to pyrolysis biochar [15, 18]. Chemical 81 characterization of GB, showing its stable structure, is well reported [4, 15, 17, 19], however studies on the effect of GB on soil and microbial processes are scarce. Concerns about the use of GB as a
soil amendment include its possible content of Polycyclic Aromatic Hydrocarbons (PAH) [20],
which proved to be highly variable, as e.g. in the studies of Wiedner [4] and Kloss [20], who
measured values up to 15 and 33 mg kg<sup>-1</sup>, respectively. Especially the wood gasification biochars
showed high PAH contents [4, 17].

The aim of this study was to evaluate the potential of the biochar residues from two gasification processes to exert a beneficial effect on soil carbon sequestration and soil quality. Through a shortterm soil incubation study and physical and chemical analyses, the objectives were to investigate if the gasification biochars: (1) contain carbon recalcitrant to microbial degradation; (2) have a potential to improve soil physical and chemical properties; (3) have any negative effects on microbial biomass and (4) have a potential for higher carbon sequestration rates than those achieved with traditional direct soil incorporation of the feedstock (i.e. straw).

#### 94 **2. Materials and methods**

#### 95 2.1. Biochar production

96 The two gasification biochars (GB) used for this study originated from continuously operated pre-97 commercial gasification demonstration plants. Straw gasification biochar (SGB) was produced in a 98 Low Temperature Circulating Fluidized Bed gasifier (LT-CFB). The straw originated from winter 99 wheat (Triticum aestivum L.) grown in Zealand, Denmark, but is of unknown provenance, date of 100 harvest and chain of custody. Commercially produced wheat straw pellets were crushed prior to LT-101 CFB gasification for optimal gasifier operation. Wood gasification biochar (WGB) was produced 102 from pine wood (Pinus spp.) chips in a TwoStage gasifier. The wood chips were commercially 103 produced with an average chip size of 50 mm, which is the optimal size for the TwoStage process, 104 and originated from Zealand, Denmark.

105 The LT-CFB gasifier (Fig. 1), developed at the Technical University of Denmark in cooperation 106 with Danish Fluid Bed Technology, is designed to gasify biomass resources with high contents of 107 low melting ash compounds (e.g. straw, manure or sewage sludge), that have proven difficult to 108 convert in other processes [1]. The process is based on separate pyrolysis and gasification fluid bed 109 reactors with a suitable circulating heating medium to transfer the heat from the gasification process 110 to the pyrolysis. The temperature is kept below the melting point of the ash components, i.e. max 111 process temperatures around 700 - 750°C. In this way, sintering of the ash and subsequent fouling 112 (from e.g. potassium) or corrosion (from e.g. chlorine) of the plant unit operations are avoided, as 113 these compounds will leave the process in solid form as ash particles.

114 Fig. 1 here.

The char conversion in the LT-CFB gasifier is a combination of sub stoichiometric oxidation of the char and steam gasification. The retention time (few seconds) in the char reactor is relatively short. The char-ash particles are though circulated in the process until they are too small/light to be separated by the primary cyclone, subsequently most of the ash and unconverted biochar is separated out of the hot gas by the secondary cyclone. The LT-CFB technology is now owned by the company Dong Energy and is being commercialized under the name Pyroneer [21].

122 The TwoStage fixed bed process (Fig. 2) was invented and developed at the Technical University of 123 Denmark and has been designed for gasification of woody biomass with low ash content [1]. The 124 TwoStage process is characterized by having pyrolysis and gasification in separate reactors with an 125 intermediate high temperature tar-cracking zone with temperatures of 1000 - 1200°C. This allows a 126 very fine control of the process temperatures, resulting in extremely low tar concentrations in the 127 produced gas, making it suitable for gas engine operation or synthesis of biofuels. Due to the high 128 temperatures, the process is only applicable for woody biomass. The char conversion is 129 predominantly a gasification reaction between carbon and steam. The char is exposed to steam at 130 high temperature, 800 - 1000°C, for a relatively long period (30+ minutes), resulting in an activated char with a high surface area. 131

132 Fig. 2 here.

133

### 134 2.2. Biochar characterization

The total content of organic C, H and O in feedstock and gasification biochar was measured on an elemental analyzer (FLASH 2000 Organic Elemental Analyzer, Thermo Scientific, Cambridge UK). The WGB and wood chips were ball milled, while the straw was ground prior to this analysis. The specific surface area was determined by the Brunauer-Emmett-Teller (BET) method by

139 nitrogen gas sorption at 77 K (Quantachrome instruments, Boynton Beach, USA). Pore size 140 distribution was obtained by Barret-Joyner-Halenda (BJH) desorption analysis after degassing the 141 samples for 2 hours at 160°C. The WGB was hand sieved in two fractions (0-0.5 and 0.5-1 mm) 142 prior to this analysis. Carbon-coated biochar samples were examined by scanning electron 143 microscope (SEM) JEOL JSM-5900 (Oxford instruments, Japan). The pH of biochar was measured 144 in a 1:5 (w/v) biochar/Milli-Q water suspension. The ash fraction was determined by heating dried 145 biochar at 550°C for 5 hours in a muffle furnace. Nine Polycyclic Aromatic Hydrocarbons (PAHs) 146 were quantified after a soxhlet extraction of 2 g sample with toluene for 48 hours by Eurofins GfA 147 (Hamburg, Germany). The measured PAHs comprised Acenaphthene, Fluorene, Phenanthrene, 148 Fluoranthene, Pyrene, Benzo(bjk)fluoranthene, Benzo(a)pyrene, Indeno(1,2,3-cd)pyrene and 149 Benzo(ghi)perylene. The particle size distribution of the biochars was determined by a vibrating 150 screen method using sieves (Retsch, Germany).

151

#### 152 **2.3.** Incubation study

153 2.3.1. Soil

A sandy loam soil from a conventional agricultural field at Bregentved estate, Zealand, Denmark (55° 22' N, 12° 05' E) was collected from the plough layer (0-25 cm), air dried and sieved to obtain a fraction  $\leq 6$  mm. The soil contained 14 % clay, 14% silt, 47 % fine sand and 24 % coarse sand. The total C content was 1.98 % and total N 0.18 %.

158 2.3.2. Experimental design

We conducted an incubation experiment including 7 treatments with 4 replicates each. In 280 ml PVC containers, 200 g soil (dry weight) were mixed thoroughly with either 2 g (1 %) or 10 g (5 %) straw or wood GB (dry weight). The treatments were: (1) Control soil without addition of organic 162 material (Control), (2) soil amended with 1 % straw (Straw1), (3) soil amended with 5 % straw 163 (Straw5), (4) soil amended with 1 % straw gasification biochar (SGB1), (5) soil amended with 5 % 164 straw gasification biochar (SGB5), (6) soil amended with 1 % wood gasification biochar (WGB1), 165 (7) soil amended with 5 % wood gasification biochar (WGB5). The straw used for this experiment was from winter wheat (Triticum aestivum L.) produced in Zealand, Denmark. After harvest, it was 166 bailed and kept dry. The straw material was ground to a particle size of  $\leq 5$  mm prior the 167 168 incubation. The water content of the soil mixtures was adjusted to 50 % of the water holding 169 capacity (determined separately for each respective mixture), and kept constant by regular weighing and watering. The containers were sealed with plastic lids with five holes (5 mm) to allow gas 170 171 exchange while minimizing moisture loss, and incubated in the dark at 22°C for 110 days. The 172 whole experiment was set up in 5 sets, enabling 5 destructive samplings. Soil respiration was 173 measured on the same set each time, which was then used for the last destructive sampling.

## 174 2.3.3. Soil analysis

Destructive soil samplings were taken at day 1, 8, 16, 32 and 110. All treatments were analyzed for 175 176 nitrate (NO<sub>3</sub>), ammonium (NH<sub>4</sub><sup>+</sup>) and dissolved organic carbon (DOC) content by extracting 10 g of fresh soil with 50 mL 0.5 mol  $K_2SO_4$  L<sup>-1</sup>. The suspensions were shaken on a horizontal shaker 177 178 for 1 h (2.5 Hz), filtrated through pleated filter paper with retention of 5-8 µm (Grade 202F, 179 Frisenette Aps, Denmark) and stored at -20°C until analysis. The extracts were analyzed for concentrations of  $NO_3^-$  and  $NH_4^+$  on an AutoAnalyzer 3 (AA3 Bran and Luebbe, Norderstedt, 180 181 Germany), and for DOC on a TOC-VCPH (Shimadzu Corp., Kyoto, Japan). The soil microbial 182 biomass carbon (SMB-C) content in each treatment was determined by vacuum incubation of 10 g soil mixture with chloroform for 24 hours, followed by K<sub>2</sub>SO<sub>4</sub> extraction. The SMB-C was 183 184 estimated from the relationship SMB-C =  $(DOC_{fumigated} - DOC_{unfumigated})/0.45$  [22]. The soil pH was 185 determined using soil-water suspension of 5 g soil and 25 ml of Milli-Q water.

#### 186 2.4. Soil respiration

The CO<sub>2</sub> emission from each sample was measured with an infra-red gas analyzer (LI-COR 8100,
Lincoln, Nebraska USA). The measuring frequency ranged from daily in the beginning of the
experiment to once a month at the end. The emissions were measured at day 1, 2, 3, 4, 5, 8, 10, 12,
15, 18, 22, 30, 36, 46, 52, 67 and 110 of the incubation period.

# 191 2.5. Statistical analysis

192 Statistical analysis of the data was performed in R, version 3.0.2. The significant interaction effect 193 between treatment and time (day) was assessed using a two-way analysis of variance (ANOVA). 194 The differences between treatments within each day of measurement were analyzed using the 195 Student-Newman-Keuls (SNK) test from the R-package "agricolae" at P $\leq$ 0.05.

#### 196 **3. Results**

#### 197 **3.1. Biochar characterization**

Table 1 illustrates that 4 and 10 % of the carbon in wood and straw feedstock, respectively, were retained in the biochar fraction. The chemical characterization of soil, feedstock and biochars is given in Table 2. Gasification of straw and wood chips led to mass loss of H and O, decrease of H/C and O/C atomic ratios and increase of ash percentage. The carbon content was higher, while H/C and O/C ratios were lower for WGB compared to SGB. The total content of 9 PAHs was 5 mg kg<sup>-1</sup> in SGB and 0.69 mg kg<sup>-1</sup> in WGB.

204 The particle size distribution of biochars is shown in Table 3. Generally, the SGB was a fine 205 powder consisting of small particles, whereas WGB was a mixture of both very small and large 206 particles (up to 1 cm). The majority of WGB-particles were larger than 0.045 mm, while the 207 opposite was true for SGB. Table 4 presents results from BET analysis. Specific surface area (SSA) 208 and pore volume were higher for WGB compared to SGB. The particle size of WGB was crucial, as 209 SSA and pore diameter were more than twice as high in particles larger than 0.5 mm compared to 210 particles smaller than 0.5 mm. SEM images illustrated in Fig. 3 show the porous structure of both 211 biochars and the higher proportion of internal pores in WGB compared to SGB.

Table 1 here.

Table 2 here.

Table 3 here.

Table 4 here.

216 Fig. 3 here.

### 217 3.2. Incubation study

#### 218 *3.2.1. Soil sampling*

The addition of straw resulted in a decrease of soil mineral nitrogen  $(N_{min})$  content  $(NO_3^- + NH_4^+)$  to almost zero already at the second sampling day and stayed at that level during the rest of the incubation period (Fig. 4). In contrast, the N<sub>min</sub> level increased over time in the control treatment and after the addition of GB. The application of the high dosage of GB resulted in about the same N<sub>min</sub> content as in the control treatment, while the low dosage of GB decreased N<sub>min</sub> significantly.

Both straw and SGB amendments caused a significantly increased content of dissolved organic carbon (DOC) in soil compared to the control treatment throughout the incubation period, except the Straw1 treatment at the last sampling day (Fig. 5A). At day 1, an especially high DOC level could be observed in the treatment with 5 % straw. On the contrary, the soil amendment with WGB led to a significantly lower DOC content compared to all other treatments throughout the incubation period.

The content of soil microbial biomass carbon (SMB-C) was - in accordance with DOC significantly increased after addition of straw compared to the rest of the treatments, especially in the beginning of the incubation (Fig. 5B). Subsequently, the SMB-C decreased until day 16 and increased again towards the end of the incubation. After 8 days of incubation, the content of SMB-C in WGB-treated soil was significantly lower than in the control treatment, and this difference became larger with time. On the contrary, there was no consistent effect of adding SGB on SMB-C: only at day 8 and 110 in the high-dosage treatment the SMB-C was lower compared to the control.

Addition of both gasification biochars increased the pH of the soil significantly, and the difference remained throughout the incubation period (Fig. 6). After 110 days, the pH increased by 1.13 and 1.36 units for SGB5 and WGB5, respectively. By contrast, soil amendment with straw significantly 240 decreased the pH in the beginning of the incubation, whereas there was no difference anymore after

241 110 days.

242 Fig. 4 here

243

244 Fig. 5 here.

245 Fig. 6 here.

246

247 *3.2.2. Soil respiration* 

248 The addition of straw to soil, at both 1 and 5%, resulted in significantly higher CO<sub>2</sub> emissions compared to control and GB treatments throughout the experimental period (Fig. 7A). The peak 249 CO<sub>2</sub> emissions in the straw and control treatments were observed during the first week of 250 251 measurement. Soil amendment with GB did not result in any initial emissions, and the treatment WGB5 even resulted in negative fluxes during the first week (Fig. 7B). After 110 days of 252 253 incubation, the cumulative total emissions were highest for straw treatments, reaching 3.51 and 9.17 mg C g<sup>-1</sup> soil emitted as CO<sub>2</sub> for Straw1 and Straw5, respectively. GB treatments resulted in 254 cumulative total emissions of  $1.7 - 2 \text{ mg C g}^{-1}$  soil emitted as CO<sub>2</sub>, slightly higher than the control 255 (1.65 mg  $g^{-1}$  soil) (data not shown). Fig. 7C illustrates the cumulative fraction of added carbon 256 257 respired within 110 days. At the end of the incubation, 78 and 41 % of straw carbon added was respired in treatments Straw1 and Straw5, respectively, while only 1-3 % of added biochar carbon 258 259 was respired.

260 Fig. 7 here.

#### 261 **4. Discussion**

#### 262 4.1. Soil carbon sequestration potential

263 A markedly smaller proportion of added carbon was respired in the GB treatments compared to the 264 straw treatments, which reflects the aromatic and recalcitrant structure of the residual carbon in 265 these biochar materials [4] after energy production during the process of gasification (Fig. 7C). The 266 addition of the high dosage of WGB resulted even in an initially negative CO<sub>2</sub> flux, probably 267 caused by binding CO<sub>2</sub> through carbonation of soluble Ca and Mg contained in the biochar, forming CaCO<sub>3</sub> and MgCO<sub>3</sub> [23, 24]. The CO<sub>2</sub> peak after straw soil incorporation was reflected in the high 268 269 initial contents of DOC and SMB-C in these treatments, confirming that the easily degradable 270 carbon pool in the straw was rapidly decomposed by the soil microbial biomass, followed by a 271 decrease in CO<sub>2</sub> emissions (Fig. 7A). The very high content of SMB-C at day 1 in the high dosage 272 straw treatment was, however, surprising (Fig. 5B), and could be attributed to chloroform-labile 273 substances in the straw itself, as also suggested by Duong [25] observing similar effects.

274 The DOC level in both biochar treatments was - in accordance with their low CO2 emissions -275 significantly lower than that in straw treatments (Fig. 5A). WGB-treated soils were even lower in 276 DOC than SGB-treated soils, which could be due to a higher content of stable carbon, probably 277 caused by higher process temperatures during the wood gasification compared to the straw 278 gasification [26]. The DOC content of SGB was higher than that of the control treatment, but did 279 not result in any corresponding CO<sub>2</sub> emissions. This might be due to CO<sub>2</sub>-binding by carbonation 280 occurring simultaneously with CO<sub>2</sub> emissions and therefore concealing soil respiration. However, 281 the DOC value in SGB treatments might also have been overestimated due to very small particles of 282 the biochar which were not retained by the filter during the extraction process. The DOC content in 283 WGB treatments was even significantly lower than in the control treatment, which might be

284 explained by a sorption of organic substances to WGB, as the SSA of wood biochar is very high 285 [14, 27]. This was also confirmed by the clear color of WGB extracts in contrast to the brownish 286 color of the other treatments. The DOC sorption by WGB could explain low CO<sub>2</sub> emissions and the 287 low content of SMB-C, as DOC is a carbon source for the microorganisms [27, 28]. However, the 288 adsorption of both DOC and microorganisms to biochar may potentially also result in higher 289 substrate consumption and therefore increase microbial activity [14]. Generally, our results confirm 290 that DOC-related parameters based on soil extraction procedures should be interpreted with caution, 291 as e.g. also Liang et al. [29] showed that the fumigation-extraction method leads to an underestimation of SMB-C in biochar-amended soil due to sorption processes. The high N 292 293 mineralization observed in the WGB treatments is another indicator that soil microbial activity was 294 not inhibited by WGB (Fig. 4). Further studies are required to assess the effect of GB on soil 295 microbial biomass.

The GB carbon stability was also confirmed by their H/C and O/C atomic ratios, that had been decreased compared to the original feedstock to values below 0.6 and 0.4, respectively (Table 2), which is in agreement with the recommended thresholds indicating carbon recalcitrance [17, 26]. The H/C and O/C atomic ratios of WGB were even lower in comparison with SGB.

### 300 4.2. Improvement of soil quality

Results from BET and SEM analyses illustrated a higher SSA and porosity in WGB compared to SGB (Table 4, Fig. 3). Besides the feedstock itself, the higher process temperature [19, 20, 27] in the wood gasification process could contribute to those characteristics, as WGB and SGB were produced at about 1000° and 700° C, respectively. However, both GBs in this study showed a relatively high SSA in comparison with other studies, where the SSA of GBs ranged from 5 to 62  $m^2 g^{-1}$  [15, 19] and that of pyrolysis biochars from 1 to 320 m<sup>2</sup> g<sup>-1</sup> [20, 27, 30]. According to

Schimmelpfennig and Glaser [17], biochar with a SSA higher than 100 m<sup>2</sup> g<sup>-1</sup> has the potential for 307 improvement of soil water and nutrient retention and porosity of the soil, which could benefit 308 309 microbes and plants. This requirement is definitely fulfilled by the WGB with an SSA of the same 310 magnitude as activated charcoal, which is probably due to the steam activation in the wood 311 gasification process [31]. The lower porosity of SGB is probably also caused by the processing, as 312 the straw fuel was pelletized and crushed, and gasified in a circulating fluidized bed (see section 313 2.1.). Cereal straw has about 6 times the amount of minerals (ash) compared to the wood chips used 314 to produce WGB, which might result in mineral matter occupying the pores of biochars or being exposed at the surface of the biochar particles and blocking the pores, thereby causing the lower 315 316 SSA [32].

Addition of both biochars resulted in an increase of soil pH due to their alkalinity (Fig. 6). The frequently described liming effect of biochar can improve plant nutrient availability, especially in case of phosphorus in low-pH soils [3, 9, 27], and may have a beneficial effect on soil fertility and plant growth on acidic soils [33].

Soil incorporation of straw with a wide C/N ratio often results in initial N immobilization [34, 35] and subsequent slow N release [11]. The N immobilization was also observed in this study in the straw treatments (Fig. 4). Contrarily, the soil application of GBs led to N levels similar to the control soil, which means that no initial adverse effects on plant growth - as they can occur after the application of pyrolysis biochar [36] - are to be expected after GB soil application. However, there is no obvious explanation for the decreased  $N_{min}$  levels compared to the control soils in the low dosage of both GBs.

The total PAH content of both biochars was well below the threshold limit of 12 mg kg<sup>-1</sup> for bioash soil application according to the Danish Ministry of the Environment (Table 2). Eventual PAH 330 content in GB originates from PAHs in the produced gas, where they are formed as a decomposition 331 product of gaseous pyrolysis tars. If the GB stays in contact with the produced gas at low 332 temperatures, PAHs may subsequently condense on the GB. Although high PAH contents are often 333 reported for wood gasification biochars [4, 17], the WGB in this study showed a value of 0.69 mg 334 kg<sup>-1</sup>, which is far below the limit, despite the high process temperatures. This is due to the 335 successful decomposition of PAHs during the TwoStage process, as the separation of the pyrolysis 336 and gasification reactors allows for a controlled gas phase partial oxidation of the pyrolysis tars 337 (Fig. 2). Consequently, the PAHs formed during the partial oxidation subsequently react with the 338 activated char in the char bed and are decomposed [37]. As a consequence of the in-process 339 decomposition, the concentration of PAHs in the produced gas is very low and hence no significant 340 PAH condensation on the WGB is possible [38]. Additionally, in the process, the WGB is separated 341 from the produced gas at high temperature (750 °C), which is significantly higher than the dew 342 point of the low PAH concentration in the gas and thus minimizes the possible condensation of 343 PAHs on the WGB.

## 344 4.3. Biomass for both energy and soil amendment

345 Biomass, such as crop residues and wood waste, is a renewable global energy source, and efficient 346 energy conversion is required to reach the ambitious political goal in many countries to obtain a 347 fossil fuel free society. According to an LCA analysis by Nguyen et al. [2], gasification is - in 348 comparison with the dominating direct combustion - more environmentally friendly due to 349 primarily three main factors: (1) a higher energy efficiency, (2) reduced emission of major air 350 pollutants and (3) a higher carbon content in the residual fraction [2]. The LT-CFB process has 351 some unique features compared to direct combustion, as it can operate on crop residues and biomass 352 related waste, which are normally problematic for direct combustion. The produced gas has a low content of ash alkali and can thus be combusted at high temperatures resulting in very efficient gas 353

utilization and energy conversion. The TwoStage gasification process allows for efficient utilization of wood at small to medium scale. By producing clean and tar free gas, which can be used in a gas engine for combined heat and power production, it is possible, even for a small scale plant, to achieve efficiencies comparable with those of large scale power plants [1].

358 Crop residue removal for energy production can potentially reduce the soil carbon and nutrient 359 content and thereby the soil quality. Powlson et al. [11] concluded that removal or incorporation of 360 straw had a small effect on soil organic carbon content; however, even a small change in SOC could 361 have large negative impacts on soil physical properties. To date, the biochar fraction extracted from 362 the gasification process is not considered a valuable product, though, if it can be developed into a soil amendment of high fertilizer and soil improver value, this will significantly improve the 363 364 economic feasibility and sustainability of the gasification technology [39]. On future markets, such 365 parameters have increasing importance, and the sustainability of a particular bioenergy chain will to 366 a large extent depend on the possibilities for its by-products recycling potential [40]. Nevertheless, 367 considering the complexity of effects of SOC on soil quality, the question, whether field application 368 of gasification biochar may replace SOC originating from crop residues, requires further research.

369 In contrast to pyrolysis, which is usually engineered to produce biochar with gas and heat as co-370 products, the main product of gasification is energy in form of syngas, while biochar is considered a 371 co-product. Thus, gasification produces more energy and less biochar compared to pyrolysis [18]. It 372 is, however, important to find a balance in the amount of carbon utilized for energy generation and 373 carbon left in the biochar for soil application. In the present study, we had a focus on both energy 374 and biochar production. In the LT-CFB process, 90 % of the feedstock-carbon was used for energy production, while 10 % remained in the biochar (Table 1). In the TwoStage process, 96 % carbon 375 376 was utilized for energy and 4 % remained in the biochar. Therefore, LT-CFB gasification of straw 377 and biochar soil amendment could on the longer term have a comparable soil carbon sequestration potential to the TwoStage gasification of wood, despite the fact that WGB carbon showed a higher stability compared to SGB. Currently, the LT-CFB gasification processes are flexible technologies, allowing an energy output of up to 97 % of the carbon input, which would reduce the SGB's carbon content from the present ca. 50 % to 20 - 30 %.

#### 383 **5. Conclusion**

384 In this study, we suggest that thermal gasification of biomass residues is able to combine the 385 production of bioenergy and a biochar fraction that can exert a positive impact on soil quality. Our 386 results showed that gasification biochar (GB) carbon is more resistant to microbial degradation 387 compared to straw carbon and has a potential for soil carbon sequestration. Furthermore, the GBs in 388 our study exhibited a potential as soil improving agents due to their high specific surface area, 389 porosity and liming effect, with PAH contents below the threshold limit. However, the differences 390 found between the two biochar materials will probably qualify them to benefit different soil 391 parameters. WGB with higher SSA, lower PAH content and higher carbon stability, caused both by 392 feedstock source but also by process conditions, could increase water holding capacity and nutrient retention on sandy soils, while SGB could be preferably used as a fertilizer or liming agent. 393 394 Gasification of straw and wood chips and field application of the biochar is therefore an integrative 395 approach combining both agriculture and forestry with the energy sector, which seems to be an 396 attractive option to maximize both energy output and soil carbon sequestration. The results of the 397 present study reveal that it is worthwhile to further test the potential of GB soil amendment, as it 398 has been done for more traditional pyrolysis biochar materials [26, 27, 34]. In this regard, it will be 399 crucial to investigate the soil application of GBs also in longer-term studies, pot and field 400 experiments, to be able to determine the effect on plant yields, soil biota and soil quality.

# 402 Acknowledgements

The financial support for this research was provided by the VILLUM Foundation. We are grateful to DONG Energy for providing us with the biochar samples. We thank Henrik Spliid for help with statistical analysis, Mette Flodgaard and Anja Nielsen for excellent technical assistance, Jakob Munkholt Christensen for help with BET analysis, Rolf Jensen for help with SEM and Esben W. Bruun for practical advice concerning LICOR measurements.

- 409
- Ahrenfeldt J, Thomsen TP, Henriksen U, Clausen LR. Biomass gasification cogeneration A 410 [1] 411 review of state of the art technology and near future perspectives. Appl Therm Eng 412 2013;50(2):1407-17. 413 Nguyen TLT, Hermansen JE, Nielsen RG. Environmental assessment of gasification [2] 414 technology for biomass conversion to energy in comparison with other alternatives: the case of wheat straw. J Clean Prod 2013;53:138-48. 415 416 [3] Müller-Stöver D, Ahrenfeldt J, Holm JK, Shalatet SGS, Henriksen U, Hauggaard-Nielsen H. Soil application of ash produced by low-temperature fluidized bed gasification: effects on 417 soil nutrient dynamics and crop response. Nutr Cycl Agroecosystems 2012;94(2-3):193-207. 418 419 [4] Wiedner K, Rumpel C, Steiner C, Pozzi A, Maas R, Glaser B. Chemical evaluation of chars 420 produced by thermochemical conversion (gasification, pyrolysis and hydrothermal 421 carbonization) of agro-industrial biomass on a commercial scale. Biomass and Bioenergy 422 2013;59:264-78. 423 Mao J-D, Johnson RL, Lehmann J, Olk DC, Neves EG, Thompson ML, et al. Abundant and [5] 424 stable char residues in soils: implications for soil fertility and carbon sequestration. Environ 425 Sci Technol 2012;46(17):9571-6. 426 Hellebrand HJ, Strähle M, Scholz V, Kern J. Soil carbon, soil nitrate, and soil emissions of [6] nitrous oxide during cultivation of energy crops. Nutr Cycl Agroecosystems 2009;87(2):175-427 428 86. 429 [7] Reeves DW. The role of soil organic matter in maintaining soil quality in continuous 430 cropping systems. Soil Tillage Res 1997;43(1-2):131-67. 431 [8] Atkinson CJ, Fitzgerald JD, Hipps N a. Potential mechanisms for achieving agricultural 432 benefits from biochar application to temperate soils: a review. Plant Soil 2010;337(1-2):1-433 18. 434 [9] Xu M, Lou Y, Sun X, Wang W, Baniyamuddin M, Zhao K. Soil organic carbon active fractions as early indicators for total carbon change under straw incorporation. Biol Fertil 435 Soils 2011;47(7):745-52. 436 437 Thomsen IK, Christensen BT. Yields of wheat and soil carbon and nitrogen contents [10] 438 following long-term incorporation of barley straw and ryegrass catch crops. Soil Use Manag 439 2004;20(4):432-8. Powlson DS, Glendining MJ, Coleman K, Whitmore AP. Implications for Soil Properties of 440 [11] 441 Removing Cereal Straw: Results from Long-Term Studies. Agron J 2011;103(1):279. González-Pérez J a, González-Vila FJ, Almendros G, Knicker H. The effect of fire on soil 442 [12] 443 organic matter--a review. Environ Int 2004;30(6):855-70.

444 Laird DA, Fleming P, Davis DD, Horton R, Wang B, Karlen DL. Impact of biochar [13] 445 amendments on the quality of a typical Midwestern agricultural soil. Geoderma 2010;158(3-446 4):443–9. 447 [14] Lehmann J, Rillig MC, Thies J, Masiello C a., Hockaday WC, Crowley D. Biochar effects on soil biota – A review. Soil Biol Biochem 2011;43(9):1812–36. 448 449 Brewer CE, Schmidt-rohr K, Satrio JA, Brown RC. Characterization of Biochar from Fast [15] 450 Pyrolysis and Gasification Systems. Environ Prog Sustain Energy 2009;28(3). Ulyett J, Sakrabani R, Kibblewhite M, Hann M. Impact of biochar addition on water 451 [16] 452 retention, nitrification and carbon dioxide evolution from two sandy loam soils. Eur J Soil 453 Sci 2014;65(1):96-104. 454 [17] Schimmelpfennig S, Glaser B. One step forward toward characterization: some important 455 material properties to distinguish biochars. J Environ Qual 2007;41(4):1001–13. 456 [18] Ahmed I, Gupta a. K. Syngas yield during pyrolysis and steam gasification of paper. Appl 457 Energy 2009;86(9):1813–21. 458 Brewer CE, Unger R, Schmidt-Rohr K, Brown RC. Criteria to Select Biochars for Field [19] 459 Studies based on Biochar Chemical Properties. BioEnergy Res 2011;4(4):312–23. 460 Kloss S, Zehetner F, Dellantonio A, Hamid R, Ottner F, Liedtke V, et al. Characterization of [20] 461 slow pyrolysis biochars: effects of feedstocks and pyrolysis temperature on biochar properties. J Environ Qual 2012;41(4):990-1000. 462 DONG Energy Power A/S [Internet]. Fredericia (DK): DONG Energy Power A/S, Pyroneer. 463 [21] 464 [Cited 2014 Aug 19] Available from: 465 http://www.dongenergy.com/da/innovation/utilising/pages/pyroneer.aspx 466 [22] Wu J, Joergensen JRG. Measurement of Soil Microbial Biomass C by Fumigation Extraction 467 - an Automated Procedure. Soil Biol Biochem 1990;22(8):0-2. (8). 468 Lim S-S, Choi W-J, Lee K-S, Ro H-M. Reduction in CO2 emission from normal and saline [23] soils amended with coal fly ash. J Soils Sediments 2012;12(9):1299-308. 469 470 [24] Ohlsson KEA. Carbonation of Wood Ash Recycled to a Forest Soil as Measured. Soil Sci 471 Soc Am J 1998:337-41. 472 Duong TTT, Baumann K, Marschner P. Frequent addition of wheat straw residues to soil [25] 473 enhances carbon mineralization rate. Soil Biol Biochem 2009;41(7):1475-82. 474 Enders A, Hanley K, Whitman T, Joseph S, Lehmann J. Characterization of biochars to [26] 475 evaluate recalcitrance and agronomic performance. Bioresour Technol 2012;114:644-53.

- 476 [27] Dai Z, Meng J, Muhammad N, Liu X, Wang H, He Y, et al. The potential feasibility for soil
  477 improvement, based on the properties of biochars pyrolyzed from different feedstocks. J
  478 Soils Sediments 2013;13(6):989–1000.
- 479 [28] Jones DL, Murphy DV, Khalid M, Ahmad W, Edwards-Jones G, DeLuca TH. Short-term
  480 biochar-induced increase in soil CO2 release is both biotically and abiotically mediated. Soil
  481 Biol Biochem 2011;43(8):1723–31.
- 482 [29] Liang B, Lehmann J, Sohi SP, Thies JE, O'Neill B, Trujillo L, et al. Black carbon affects the
  483 cycling of non-black carbon in soil. Org Geochem 2010;41(2):206–13.
- 484 [30] Sun Z, Moldrup P, Elsgaard L, Arthur E, Bruun EW, Hauggaard-Nielsen H, et al. Direct and
  485 Indirect Short-term Effects of Biochar on Physical Characteristics of an Arable Sandy Loam.
  486 Soil Sci 2013;178(9):465–73.
- 487 [31] Anderson N, Jones J, Page-Dumroese D, McCollum D, Baker S, Loeffler D, et al. A
  488 Comparison of Producer Gas, Biochar, and Activated Carbon from Two Distributed Scale
  489 Thermochemical Conversion Systems Used to Process Forest Biomass. Energies
  490 2013;6(1):164–83.
- 491 [32] Downie A, Crosky A, Munroe P. Physical properties of biochar. In: Lehmann J, Joseph S,
  492 editors. Biochar Environmental Management: Science and Technology. London: Earthscan;
  493 2009. p. 13–32.
- 494 [33] Deal C, Brewer CE, Brown RC, Okure M a. E, Amoding A. Comparison of kiln-derived and
   495 gasifier-derived biochars as soil amendments in the humid tropics. Biomass and Bioenergy
   496 2012;37:161–8.
- 497 [34] Bruun EW, Ambus P, Egsgaard H, Hauggaard-Nielsen H. Effects of slow and fast pyrolysis
  498 biochar on soil C and N turnover dynamics. Soil Biol Biochem 2012;46:73–9.
- [35] Zavalloni C, Alberti G, Biasiol S, Vedove GD, Fornasier F, Liu J, et al. Microbial
   mineralization of biochar and wheat straw mixture in soil: A short-term study. Appl Soil Ecol
   2011;50:45–51.
- [36] Nelissen V, Ruysschaert G, Müller-Stöver D, Bodé S, Cook J, Ronsse F, et al. Short-Term
   Effect of Feedstock and Pyrolysis Temperature on Biochar Characteristics, Soil and Crop
   Response in Temperate Soils. Agronomy 2014;4(1):52–73.
- Egsgaard H, Ahrenfeldt J, Ambus P, Schaumburg K, Henriksen UB. Gas cleaning with hot
   char beds studied by stable isotopes. J Anal Appl Pyrolysis 2014;107:174–82.
- 507 [38] Henriksen U, Ahrenfeldt J, Jensen TK, Gøbel B, Bentzen JD, Hindsgaul C, et al. The design,
  508 construction and operation of a 75kW two-stage gasifier. Energy 2006;31(10-11):1542–53.
- 509 [39] Blanco-Canqui H. Crop Residue Removal for Bioenergy Reduces Soil Carbon Pools: How
   510 Can We Offset Carbon Losses? BioEnergy Res 2012;6(1):358–71.

- Taheripour F, Hertel TW, Tyner WE, Beckman JF, Birur DK. Biofuels and their by-products: Global economic and environmental implications. Biomass and Bioenergy 2010;34(3):278– [40]
- 89.

515 Figure Captions:

516 Fig. 1 – Schematic of Low Temperature Circulating Fluidized Bed gasifier (LT-CFB) [21].

517 Fig. 2 – Schematic of the TwoStage gasifier [1].

518 Fig. 3 - Scanning electron microscope (SEM) images; left: straw gasification biochar (SGF) and 519 right: wood gasification biochar (WGB).

Fig. 4 – Content of soil mineral nitrogen ( $N_{min}$ ) during the incubation period of 110 days. Straw1= soil amended with 1% straw, Straw5= soil amended with 5 % straw, SGB1= soil amended with 1 % straw gasification biochar, SGB5= soil amended with 5 % straw gasification biochar, WGB1= soil amended with 1 % wood gasification biochar, WGB5= soil amended with 5 % wood gasification biochar, Control= untreated soil. Values presented are means with standard error bars (n = 4). Treatments with different letters are significantly different at the last day of the incubation (P < 0.05).

Fig. 5 – A) Content of dissolved organic carbon (DOC) in soil during the incubation period of 110 days. B) Content of soil microbial biomass-carbon (SMB-C) in soil during the incubation period of 110 days. For treatment abbreviations, see Fig. 4. Values presented are means with standard error bars (n = 4). Treatments with different letters are significantly different at the last day of the incubation (P < 0.05).

Fig. 6 – Soil pH at day 1, 8, and 110 of the incubation period. For treatment abbreviations, see Fig. 4. Values presented are means with standard error bars (n = 3). Treatments with different letters are significantly different (P < 0.05).

Fig. 7 – A)  $CO_2$  fluxes from soil during the incubation period of 110 days. B)  $CO_2$  fluxes during the first 8 days of incubation. C) Cumulative fraction of added carbon respired from soil during the 537 incubation period of 110 days. For treatment abbreviations, see Fig. 4. Values presented are means 538 with standard error bars (n = 4). Treatments with different letters are significantly different at the 539 last day of the incubation (P < 0.05).

	Percentage TwoStage input		Percentage LT-CFB input	
-	Fractional distribution			
	Carbon (%)	Energy (%)	Carbon (%)	Energy (%)
Biomass feedstock	100	100	100	100
Product gas output	96	92	90	85-87
Biochar output	4	4	10	10
Loss	-	4	-	3-5

Table 1 – Carbon and energy balance for TwoStage gasifier and Low-temperature circulating fluidized bed gasifier (LT-FCB) reflecting the carbon loss in the GB used in this study.

	Soil	Straw	Wood chips	SGB	WGB
C (%)	1.98	45.50	52.04	46.80	65.29
H (%)	-	5.52	7	0.97	0.63
O (%)	-	36.85	41.16	13.11	8.99
H/C atomic ratio	-	1.46	1.61	0.25	0.12
O/C atomic ratio	-	0.61	0.59	0.21	0.10
pH (water)	7.9	-	-	11.6	11.1
Ash (%)	-	4.85	0.75	52	33
$\Sigma PAH^{a} (mg kg^{-1})$	-	-	-	5	0.69

Table 2 - Chemical characterization of soil, feedstock and biochars (SGB = straw gasification biochar, WGB = wood gasification biochar).

<sup>a</sup> Sum af Acenaphthene, Fluorene, Phenanthrene, Fluoranthene, Pyrene, Benzo(bjk)fluoranthene, Benzo(a)pyrene, Indeno(1,2,3-cd)pyrene and Benzo(ghi)perylene.

# Table3

Biochar	Particle size distribution in % of dry mass			
	< 0.045 mm	0.045-0.125 mm	>0.125 mm	
SGB	89.4	10.3	0.3	
WGB	33.0	13.7	53.3	

Table 3 - Particle size distribution of straw gasification biochar (SGB) and wood gasification biochar (WGB).

10010
-------

Biochar	Particle size (mm)	$SSA (m^2 g^{-1})$	Pore volume (cm <sup>3</sup> g <sup>-1</sup> )	Pore diameter (nm)
SGB	0-1	75	0.04	3.71
WGB	0-0.5	426	0.52	1.43
WGB	0.5-1	1027	0.58	3.73

Table 4: BET specific surface area (SSA), pore volume and diameter of straw gasification biochar (SGB) and wood gasification biochar (WGB).



All tar concentrations measured per volume gas produced.

All gas volumes measured at Normal Temperature and Pressure i.e. 20 °C and 101325 kg m<sup>-1</sup> s<sup>-2</sup>







Figure Captions:

Fig. 1 – Schematic of Low Temperature Circulating Fluidized Bed gasifier (LT-CFB) [21].

Fig. 2 – Schematic of the TwoStage gasifier [1].

Fig. 3 - Scanning electron microscope (SEM) images; left: straw gasification biochar (SGF) and right: wood gasification biochar (WGB).

Fig. 4 – Content of soil mineral nitrogen ( $N_{min}$ ) during the incubation period of 110 days. Straw1= soil amended with 1% straw, Straw5= soil amended with 5% straw, SGB1= soil amended with 1% straw gasification biochar, SGB5= soil amended with 5% straw gasification biochar, WGB1= soil amended with 1% wood gasification biochar, WGB5= soil amended with 5% wood gasification biochar, Control= untreated soil. Values presented are means with standard error bars (n = 4). Treatments with different letters are significantly different at the last day of the incubation (P < 0.05).

Fig. 5 – A) Content of dissolved organic carbon (DOC) in soil during the incubation period of 110 days. B) Content of soil microbial biomass-carbon (SMB-C) in soil during the incubation period of 110 days. For treatment abbreviations, see Fig. 4. Values presented are means with standard error bars (n =4). Treatments with different letters are significantly different at the last day of the incubation (P < 0.05).

Fig. 6 – Soil pH at day 1, 8, and 110 of the incubation period. For treatment abbreviations, see Fig. 4. Values presented are means with standard error bars (n = 3). Treatments with different letters are significantly different (P < 0.05).

Fig. 7 – A) Carbon emitted as CO<sub>2</sub> from soil during the incubation period of 110 days. B) Carbon emitted as CO<sub>2</sub> during the first 8 days of incubation. C) Cumulative fraction of added carbon respired from soil during the incubation period of 110 days. For treatment abbreviations, see Fig. 4. Values presented are means with standard error bars (n = 4). Treatments with different letters are significantly different at the last day of the incubation (P < 0.05).





